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Attorney Docket No. SALK1510-3

\_\_\_ NEW PATENT APPLICATION
X CONTINUATION-IN-PART

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Sir:

Transmitted herewith for filing is the new patent application of

Inventors: Ronald M. Evans, J. Don Chen and Peter Ordentlich

For: A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR

This is a request for filing a continuation-in-part under 35 U.S.C. 111(A) and 37 C.F.R. 1.53(b), of U.S. Application Serial No. 08/522,726, filed September 1, 1995, now pending.

Enclosed are:

- X 75 pages of the Specification, which includes 7 pages of the claims and 1 page of the Abstract;
- X 12 sheets of drawing(s) \_\_\_ Formal; X Informal;
- X A Declaration (unexecuted);
- X 67-Page Sequence Listing;
- X computer readable disk containing Sequence Listing; and
- X Statement Under 37 C.F.R. §§1.821(f) and (g).

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In re Application of: Evans et al.

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The filing fee has been calculated as shown below:

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Respectfully submitted,

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# APPLICATION

For

# UNITED STATES LETTERS PATENT

on

# A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR

by

Ronald M. Evans, J. Don Chen and Peter Ordentlich

Sheets of Drawings: Twelve (12)

Docket No.: SALK 1510-3

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# A Family of Transcriptional Co-repressors that Interact with Nuclear Hormone Receptors and Uses Therefor

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#### **RELATED APPLICATIONS**

This application is a continuation-in-part application of pending United States application Serial No. 08/522,726, filed September 1, 1995 and is related to United States application Serial No. \_\_\_\_\_\_\_, filed on even date herewith, each of which is incorporated herein in its entirety by reference.

### FIELD OF THE INVENTION

The present invention relates to intracellular receptors, methods for the modulation thereof, and methods for the identification of novel ligands therefor. In a particular aspect, the present invention relates to methods for the identification of compounds which function as ligands (or ligand precursors) for intracellular receptors. In another aspect, the present invention relates to novel chimeric constructs and uses therefor.

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#### **BACKGROUND OF THE INVENTION**

A central problem in eukaryotic molecular biology continues to be the elucidation of molecules and mechanisms that mediate specific gene regulation. As part of the scientific attack on this problem, a great deal of work has been done in efforts to identify ligands (i.e., exogenous inducers) which are capable of mediating specific gene regulation. Additional work has been done in efforts to identify other molecules involved in specific gene regulation.

Although much remains to be learned about the specifics of gene regulation, it is known that ligands modulate gene transcription by acting in concert with intracellular components, including intracellular receptors and discrete DNA sequences known as hormone response elements (HREs).

The identification of compounds that directly or indirectly interact with intracellular receptors, and thereby affect transcription of hormone-responsive genes, would be of significant value, e.g., for therapeutic applications.

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Transcriptional silencing mediated by nuclear receptors plays an important role in development, cell differentiation, and is directly linked to the oncogenic activity of v-erbA. The mechanism underlying this effect is unknown but is one key to understanding the molecular basis of hormone action. Accordingly, the identification of components involved in transcriptional silencing would represent a great advance in current understanding of mechanisms that mediate specific gene regulation.

Other information helpful in the understanding and practice of the present invention can be found in commonly assigned United States Patent Nos. 5,071,773, 4,981,784, 5,260,432, and 5,091,513, all of which are hereby incorporated herein by reference in their entirety.

### BRIEF DESCRIPTION OF THE INVENTION

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The present invention overcomes many problems in the art by providing a family of receptor interacting co-repressors, referred to herein as "SMRT co-repressor", i.e., a silencing mediator (co-repressor) for retinoic acid receptor (RAR) and thyroid hormone receptor (TR). *In vivo*, members of the SMRT family of co-repressors function as potent co-repressors. A GAL4 DNA binding domain (DBD) fusion with a SMRT co-repressor behaves as a frank repressor of a GAL4-dependent reporter.

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Together, these observations identify a novel family of cofactors that is believed to represent an important mediator of hormone action.

Accordingly, the present invention provides isolated silencing mediators of retinoic acid and thyroid hormone receptors, and isoforms or peptide portions thereof (SMRT co-repressors), that modulate transcriptional potential of members of the nuclear receptor superfamily. Such SMRT co-repressors comprise a repression domain having less than about 83% identity with a Sin3A interaction domain of N-CoR (amino acids 255 to 312 of SEQ ID NO: 11); less than about 57% identity with repression domain 1 of N-CoR (amino acids 1 to 312 of SEQ ID NO: 11); less than about 66% identity with a SANT domain of N-CoR (amino acids 312 to 668 of SEQ ID NO: 11) and/or; less than about 30% identity with repression domain 2 of N-CoR (amino acids 736 to 1031 of SEQ ID NO: 11).

In accordance with yet another embodiment of the present invention, there are provided isolated peptides comprising at least a portion of the invention SMRT co-repressor six contiguous amino acids of an amino acid sequence selected from the group consisting of:

amino acids 1 to 1030 of SEQ ID NO: 5;
amino acids 1 to 1029 of SEQ ID NO: 7;
amino acids 1 to 809 of SEQ ID NO: 9;
and conservative variations thereof,
provided the peptide is not identical to a sequence of SEQ ID NO: 11.

In addition, there are provided isolated antibodies that bind specifically to invention isolated peptides. There are also provided chimeric molecules comprising invention isolated peptides and at least a second molecule. Also provided are complexes comprising an invention SMRT co-repressor and a member of the superfamily of nuclear receptors and isolated antibodies that bind to such complexes.

Accordingly, the present invention provides isolated polynucleotides encoding members of the newly described family of silencing mediators of retinoic acid and thyroid hormone receptor or an isoform or peptide portion thereof (SMRT co-repressor), or an isolated polynucleotide complementary thereto. In addition, there are provided vectors comprising invention polynucleotides, as well as host cells containing invention polynucleotides.

In additional embodiments of the present invention, there are provided methods for identifying agents that modulate the repressor potential of a SMRT corepressor.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates a function of an invention SMRT co-repressor.

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In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell.

In another embodiment according to the present invention, there are provided methods of identifying a molecule that interacts specifically with a SMRT co-repressor.

#### BRIEF DESCRIPTION OF THE FIGURES

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Figure 1 shows the quantitation by phosphoimager of a dose-dependent dissociation of SMRT from RAR or TR by all-*trans* retinoic acid (atRA) or thyroid hormone (triiodothyronine or T3).

Figure 2 presents amino acid (aa) sequences of SMRT (Genbank accession number XXXXX). The aa sequence presented in parentheses (i.e., residues

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1330-1376) is an alternatively spliced insert which is not present in the original two-hybrid clone (C-SMRT, aa 981 to C-terminal end). The proline-rich N-terminal domain (aa 1-160) and the glutamine-rich region (aa 1061-1132), as well as the ERDR and SG regions, are also indicated. The C-terminal region of SMRT (aa 1201 to C-terminal end) shows 48% aa identity to RIP13 (Seol et al., *Molecular Endocrinology* 9:72-85 (1995)). The rest of the sequence of RIP13 shows 22% aa identity to SMRT (aa 819-1200).

Figure 3 illustrates mediation of the silencing effect of hRAR $\alpha$  and hTR $\beta$  by SMRT in vivo.

Figure 3(A) illustrates that v-erbA reverses the silencing effect of GAL-RAR (GAL4 DBD-hRARa 156-462) while SMRT restores the silencing effect.

Figure 3(B) illustrates that the RAR403 truncation mutant reverses the silencing effect of GAL-TR (GAL4 DBD-hTRβ 173-456) while SMRT restores the silencing effect.

Figure 3(C) illustrates that v-erbA and full length SMRT or C-SMRT 20 have no effect on GAL-VP16 activity.

Figure 3(D) illustrates that a GAL4 DBD fusion of full length SMRT represses the thymidine kinase basal promoter activity containing four GAL4 binding sites. The fold of repression was calculated by dividing the normalized luciferase activity transfected with the GAL4 DBD alone by those transfected with indicated amount of GAL DBD fusion constructs.

Figure 4 provides an alignment of the human SMRT (SEQ ID NO: 5) and mouse SMRTα (SEQ ID NO: 7) amino acid sequences. Proteins were aligned using the CLUSTAL alignment program. Underlined sequence of mouse SMRTα corresponds to the amino acid sequences that are deleted in mouse SMRTβ. The

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arrow indicates the start point of the previously described human SMRT co-repressor (sSMRT).

Figures 5A and 5B provide alignments of the human SMRT and human N-CoR co-repressors.

Figure 6A is a graph showing the results of transactivation experiments using transcripts encoding a detectable reporter and either wild type EcR (Ecr wt), a repression-Defective EcR allele Ecraa<sup>483T</sup> (EcRA483T) or vp16 activation domain fused to Ultraspiracle (vp16-USP).

Figure 6B is a graph showing the results of transactivation experiments using CMV promoter-driven expression vectors. Wild-type EcR or EcR A483T was cotransfected with vp16-USP and Gal4-c-SMRT (aa 981 to C terminus) (Chen and Evans, *Nature* 377:454-457, (1995)) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element.

Figure 6C shows alignment of EcR, rTR, hRAR, and rRev-erbA receptor sequences and the secondary structure in the LBD signature motif region. Conserved residues are marked in dark. The mutation 483 (AT) is marked at the top of the corresponding residue.

Figure 7 is a graph showing β-galactosidase activity in a yeast two-hybrid screen with pAS-EcR as bait. pAS-EcR is a fusion gene with the region corresponding to aa 223-878 of EcRB1 fused C-terminally to the Gal4-DBD of the pAS1-CYH2 construct (Durfee et al., *Genes Dev* 7:555-569 (1993)); other Gal4-DBD-based nuclear receptor constructs used in this yeast two-hybrid assay include: USP (aa 50-508), hRAR (aa 186-462) and hTR (aa 121-410) (Schulman et al., *Proc. Natl. Acad. Sci. USA*, 92:8288-8292, (1995)), and SMRT (Chen and Evans, (1995), *supra*).

 $\beta$ -galactosidase activities were quantified by liquid assay for yeast cells treated either without ligand or with 3  $\mu$ M of corresponding hormone. All-trans retinoic acid (ATRA) is a ligand of RAR; 3,3',5-triiodothyroacetic acid (T3) is a ligand of TR. RAR, retinoic acid receptor; TR, thyroid hormone receptor.

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Figure 8A shows the complete amino acid sequence of the SMRTER protein (SEQ ID NO: 12). The underlined regions represent the residues also conserved in SMRT and N-CoR. The gray box indicates the sequences of the E52 clone.

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Figure 8B is a schematic structural diagram of SMRTER, SMRT, and N-CoR showing the conserved SNOR, SANT, GST, ITS, D/ER repeat, and LSD motifs with their designated patterns positioned in their relative regions in each protein.

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Figure 9. Sequence Comparison of SMRTER, SMRT, N-CoR, and Other Related Proteins. The SANT domains of various proteins are listed. Percent identities/similarities compared to SMRTER are shown on the right. Two potential helices are predicted in the N-terminal half of the SANT domain. Black boxes indicate identical sequences; gray boxes, similar or partially identical sequences.

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Figure 10 is a schematic representation showing functional domains in SMRTER. Numbers on the left represent the regions in SMRTER used to generate the Gal4-DBD fusion genes. Black stippled bars indicate the locations of EcRinteracting domains; gray stippled bars indicate repression domains. Plus signs indicate that a positive interaction between SMRTER and the EcR complex and repression of basal activity by Gal4-SMRTER is significant. ERID = ecdysone receptor-interacting domain; SMRD = SMRTER repressor domain.

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Figure 11A is a graph showing the interaction of ERID1 AND ERID2 with the EcR complex. Figure 11B is a graph showing the results of competition

between ERID1, ERID2 and c-SMRT for binding to EcR. Figure 11C is a graph showing that EcR A483T disrupts the interaction with ERID1 and ERID2.

Figure 12A shows the results of mapping three repression domains. To examine repressive activity, transcriptional activity of each Gal4-SMRTER fusion was compared to the basal activity of Gal4-DBD on reporter. Only repression with value approximately 5-fold or over is considered positive (+).

Figure 12B is a schematic representation of mapping the SMRTER-interacting domain in mSin3A and dSin3A. Yeast two-hybrid assays were used to assess the interaction between each Gal4-DBD-based fusion gene of each SMRD and the ACT-based fusion genes of mSin3A and dSin3A. The numbers indicate the region in either mSin3A or in dSin3A used to generate the ACT fusion genes. Constructs of mSin3A were described previously in Nagy et al., *Cell* 89:373-380, (1997).

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Figure 12C shows an alignment of SMRD3 of SMRTER and an mSin3-interacting domain of N-CoR. Conserved residues are boxed in gray. An asterisk indicates the region where the mutation (Gly) was generated. Minus signs indicate that the interaction between SMRD3 and Sin3A was not detectable in the yeast two-hybrid assays. Repression was measured by comparing the transcriptional activity of Gal4-SMRD3 M2 or Gal4-SMRD3 M3 to that of wild-type Gal4-SMRD3 using transfection experiments as described above.

# **DETAILED DESCRIPTION OF THE INVENTION**

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In accordance with the present invention, there is provided a family of isolated SMRT co-repressors, and isoforms and peptide portions thereof, that modulate transcriptional potential of members of the nuclear receptor superfamily. Exemplary members of this family are co-repressors having substantially the same sequence as residues 1-1329 plus 1376-1495, as set forth in SEQ ID NO:1, optionally further

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comprising the amino acid residues set forth in SEQ ID NO:2 (i. e., residues 1330-1375 of SEQ ID NO:1).

In another embodiment according to the present invention, the invention SMRT co-repressor comprises a repression domain having less than about 83% identity with a Sin3A interaction domain of N-CoR (as amino acids 255 to 312 of SEQ ID NO: 11); less than about 57% identity with repression domain 1 of N-CoR (amino acids 1 to 312 of SEQ ID NO: 11); less than about 66% identity with a SANT domain of N-CoR (amino acids 312 to 668 of SEQ ID NO: 11 and/or; less than about 30% identity with repression domain 2 of N-CoR (amino acids 736 to 1031 of SEQ ID NO: 11). Such an encoded SMRT co-repressor or peptide portion thereof is further characterized in that it can modulate transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor).

The invention SMRT co-repressors are additionally exemplified by a full length human SMRT co-repressor, (amino acids 1 to 2517 of SEQ ID NO: 5); and by two mouse SMRT isoforms, including a longer SMRT isoform designated mouse SMRT $\alpha$ , which has an amino acid sequence set forth as amino acids 1 to 2473 of SEQ ID NO: 7; and a shorter SMRT isoform designated mouse SMRT $\beta$  (amino acids 1 to 2253 of SEQ ID NO: 9). As compared to the mouse SMRT $\alpha$  isoform (SEQ ID NO: 7), the mouse SMRT $\beta$  isoform (SEQ ID NO: 9) has a deletion corresponding to amino acids 36 to 254 of SEQ ID NO: 7.

A peptide portion of a SMRT co-repressor is exemplified herein by
amino acids 1 to 1031 of SEQ ID NO: 5; amino acids 1 to 1031 of SEQ ID NO: 7;
and amino acids 1 to 813 of SEQ ID NO: 9, which includes the entire amino terminal
domain of a SMRT co-repressor. Additional peptide portions of a SMRT corepressor are exemplified by amino acids 1 to 303 of SEQ ID NO: 7; amino acids 845
to 986 of SEQ ID NO: 7; amino acids 427 to 663 of SEQ ID NO: 7; amino acids 845
to 1055 of SEQ ID NO: 7; amino acids 736 to 1031 of SEQ ID NO: 7; and amino
acids 1 to 85 of SEQ ID NO: 9, which are sub-domains of the amino terminal domain

of mouse SMRTα that have nuclear receptor repressor potential, as well as by the corresponding peptide portions of human SMRT and corresponding peptide portions of mouse SMRTβ, which can modulate the transcriptional potential of a nuclear receptor, particularly a nuclear receptor that is in the form of a dimer, for example, a thyroid hormone receptor homodimer, a retinoic acid receptor homodimer, a retinoid X receptor homodimer, a thyroid hormone receptor-retinoid X receptor heterodimer, or a retinoic acid receptor-retinoid X receptor heterodimer. In addition, the invention relates to isolated peptides that contain at least six contiguous amino acids of an amino acid sequence set forth as amino acids 1 to 1030 of SEQ ID NO: 5; amino acids 1 to 1029 of SEQ ID NO: 5; or amino acids 1 to 809 of SEQ ID NO: 9, provided the SMRT peptide is not identical to a sequence of N-CoR (SEQ ID NO: 11).

Invention co-repressor can be an invertebrate SMRT co-repressor, such as the Drosophilia SMRTER co-repressor having an amino acid sequence as set forth in SEQ ID NO: 12, or conservative variations thereof.

Additional exemplary co-repressors are those containing one or both of the receptor interacting domains (ERID1 and ERID2) identified in the Drosophilia co-repressor. For example, co-repressors containing such receptor interacting domains can be selected from the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 1698-1924 of SEQ. ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, amino acids 1698-2063 of SEQ. ID NO:12, amino acids 2094-3040 of SEQ. ID NO:12, amino acids 2929-3181 of SEQ. ID NO:12, amino acids 542-950 of SEQ. ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2929-3040 of SEQ ID NO:12, and

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thereof.

amino acids 2951-3038 of SEQ ID NO:12, and conservative variations thereof.

Additional exemplary co-repressors are those containing one or more of three autonomous repressor domains termed SMRD1, SMRD2, and SMRD3 identified in the SMRTER co-repressor. For example, invention co-repressors can contain the following autonomous repressor domains derived from Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 542-950 of SEQ. ID NO:12 amino acids 1698-1924 of SEQ ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, and conservative variations

Conservative variations of the above-described SMRT co-repressors

are also contemplated to be within the scope of the present invention. Moreover,
proteins, polypeptides and peptides having at least 80% sequence identity with any of
the SMRT co-repressors described herein are also contemplated to be within the scope
of the invention.

In another embodiment according to the present invention, there are provided chimeric molecules comprising invention isolated peptides and at least a second molecule. For example, the second molecule in invention chimeric molecule can be a polynucleotide or a polypeptide. In one embodiment, the chimeric molecule is a fusion polypeptide comprising a SMRT co-repressor operably linked to a DNA binding domain of a transcription factor.

In another embodiment according to the present invention, there are provided isolated antibodies that bind specifically to invention isolated peptides. In one embodiment, an antibody of the invention binds specifically to an epitope of a SMRT co-repressor. Such an antibody is characterized, in part, in that it does not substantially crossreact with an N-CoR polypeptide. In another embodiment, an

antibody of the invention binds specifically to a complex, which includes a SMRT corepressor or peptide portion thereof of the invention, a nuclear receptor and, optionally, a DNA regulatory element that is specifically bound by the nuclear receptor. Such an antibody is characterized, in part, in that it does not substantially crossreact with the nuclear receptor, either alone or bound to the DNA regulatory element. An antibody of the invention can be a monoclonal antibody, or can be one of a plurality of polyclonal antibodies, which essentially is a mixed population of monoclonal antibodies. The invention also relates to a cell line, which produces the monoclonal antibody of the invention.

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Such antibodies can be employed for a variety of purposes, e.g., for studying tissue localization of invention SMRT co-repressor, the structure of functional domains, the purification of receptors, as well as in diagnostic applications, therapeutic applications, and the like. Preferably, for therapeutic applications, the antibodies employed will be monoclonal antibodies.

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The above-described antibodies can be prepared employing standard techniques, as are well known to those of skill in the art, using the invention SMRT corepressor or portions thereof as antigens for antibody production. Both anti-peptide and anti-fusion protein antibodies can be used [see, for example, Bahouth et al. (1991) Trends Pharmacol Sci. vol. 12:338-343; Current Protocols in Molecular Biology (Ausubel et al., eds.) John Wiley and Sons, New York (1989). Factors to consider in selecting portions of invention SMRT co-repressor for use as immunogen (as either a synthetic peptide or a recombinantly produced bacterial fusion protein) include antigenicity, accessibility (i.e., where the selected portion is derived from, e.g., the ligand binding domain, DNA binding domain, dimerization domain, and the like), uniqueness of the particular portion selected (relative to known receptors and co-repressors therefor), and the like.

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In another embodiment according to the present invention, there are provided complexes comprising an invention SMRT co-repressor and a member of

the nuclear receptor superfamily and isolated antibodies that bind to such complexes. The nuclear receptor can be in the form of a monomer or dimer, for example, a thyroid hormone receptor homodimer, a retinoic acid receptor homodimer, a retinoid X receptor homodimer, a thyroid hormone receptor-retinoid X receptor heterodimer, a retinoic acid receptor-retinoid X receptor heterodimer, a ecdysone receptor-Ultraspiracle receptor heterodimer, and the like. Optionally or alternatively, the complex can include a DNA regulatory element, bound specifically by a DNA binding domain of the nuclear receptor.

The above-described complexes optionally further comprise a response element for the member of the nuclear receptor superfamily. Such response elements are well known in the art. Thus, for example, RAR response elements are composed of at least one direct repeat of two or more half sites separated by a spacer of five nucleotides. The spacer nucleotides can independently be selected from any one of A, C, G or T.

Each half site of response elements contemplated for use in the practice of the invention comprises the sequence

#### -RGBNNM-,

#### wherein

R is selected from A or G;
B is selected from G, C, or T;
each N is independently selected from A, T, C, or G; and
M is selected from A or C;

with the proviso that at least 4 nucleotides of said -RGBNNM- sequence are identical with the nucleotides at corresponding positions of the sequence -AGGTCA-. Response elements employed in the practice of the present invention can optionally be preceded by N<sub>x</sub>, wherein x falls in the range of 0 up to 5.

Similarly, TR response elements can be composed of the same half site repeats, with a spacer of four nucleotides. Alternatively, palindromic constructs as have been described in the art are also functional as TR response elements.

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The above-described SMRT co-repressor/dimeric receptor complexes can be dissociated by contacting the complex with a ligand for the member of the nuclear receptor superfamily.

As employed herein, the term "ligand (or ligand precursor) for a member of the nuclear receptor superfamily" (i.e., intracellular receptor) refers to a substance or compound which, in its unmodified form (or after conversion to its "active" form), inside a cell, binds to receptor protein, thereby creating a ligand/receptor complex, which in turn can activate an appropriate hormone response element. A ligand therefore is a compound which acts to modulate gene transcription for a gene maintained under the control of a hormone response element, and includes compounds such as hormones, growth substances, non-hormone compounds that modulate growth, and the like. Ligands include steroid or steroid-like hormone, retinoids, thyroid hormones, pharmaceutically active compounds, and the like. Individual ligands may have the ability to bind to multiple receptors.

Accordingly, as employed herein, "putative ligand" (also referred to as "test compound") refers to compounds such as steroid or steroid-like hormones, pharmaceutically active compounds, and the like, that are suspected to have the ability to bind to the receptor of interest, and to modulate transcription of genes maintained under the control of response elements recognized by such receptor.

In another embodiment according to the present invention, there are provided polynucleotides encoding members of the above-described family of silencing mediators of retinoic acid and thyroid hormone receptor, or an isoform or peptide portion thereof (SMRT co-repressors), or an isolated polynucleotide complementary thereto.

Invention polynucleotides include those encoding a SMRT corepressor comprises a repression domain having

- a) less than about 83% identity with a Sin3A interaction domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
- b) less than about 57% identity with repression domain 1 of N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11;
- c) less than about 66% identity with a SANT domain of N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
- d) less than about 30% identity with repression domain 2 of N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11.

In addition, an invention polynucleotide can encode a mouse SMRTβ isoform having an amino acid sequence as set forth in SEQ ID NO: 9 or conservative variations thereof, or a polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 8.

Further examples of invention polynucleotides are those comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 6; nucleotides 1 to 2801 of SEQ ID NO: 8; nucleotides 1 to 8388 of SEQ ID NO: 6; nucleotides 1 to 7465 of SEQ ID NO: 8; and nucleotides 1 to 8561 of SEQ ID NO: 4.

The invention polynucleotides further comprise those encoding a human SMRT co-repressor having an amino acid sequence as set forth in SEQ ID NO: 5, for example, a nucleotide sequence as set forth in SEQ ID NO: 4; by a polynucleotide encoding a mouse SMRTα isoform having an amino acid sequence as set forth in SEQ ID NO: 7, for example, a nucleotide sequence as set forth in SEQ ID NO: 6; and by a polynucleotide encoding a mouse SMRTβ isoform having an amino acid sequence as set forth in SEQ ID NO: 9, for example, a nucleotide sequence as set forth in SEQ ID NO: 8. A polynucleotide of the invention is further exemplified by

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polynucleotides encoding peptide portions of a SMRT co-repressor such as a polynucleotide containing nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 7; or nucleotides 1 to 2801 of SEQ ID NO: 8, which can repress the transcriptional activity of nuclear receptor, particularly a nuclear receptor that is in the form of dimer.

Additional invention polynucleotides include those encoding a full length insect SMRTER co-repressor having an amino acid sequence as set forth in SEQ ID NO: 12, or conservative variations thereof.

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Additional exemplary invention polynucleotides are those encoding one or both of the receptor interacting domains (ERID1 and ERID2) identified in invention co-repressors. For example, polynucleotides encoding such receptor interacting domains can be selected from those encoding the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 1698-1924 of SEQ. ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, amino acids 1698-2063 of SEQ. ID NO:12, amino acids 2094-3040 of SEQ. ID NO:12, amino acids 2929-3181 of SEQ. ID NO:12, amino acids 542-950 of SEQ. ID NO:12, amino acids 2094-3181 of SEQ ID NO:12, amino acids 2929-3040 of SEQ ID NO:12, amino acids 2929-3040 of SEQ ID NO:12, and amino acids 2951-3038 of SEQ ID NO:12,

and conservative variations thereof.

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Additional exemplary invention polynucleotides are those encoding one or more of three autonomous repressor domains termed SMRD1, SMRD2, and SMRD3 identified in the invention co-repressors. For example, polynucleotides encoding such autonomous repressor domains can be selected from those encoding

the following segments of the Drosophilia SMRTER co-repressor (SEQ. ID 12):

amino acids 542-950 of SEQ. ID NO:12 amino acids 1698-1924 of SEQ ID NO:12, amino acids 2951-3038 of SEQ. ID NO:12, and conservative variations

thereof.

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A polynucleotide that has at least 80% sequence identity or that hybridizes, (preferably under high stringency conditions) with any one of the above-described polynucleotides is also contemplated to be within the scope of this invention.

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A polynucleotide of the invention can be operably linked to a second nucleotide sequence and, therefore, can encode a fusion polypeptide, for example, a SMRT co-repressor, or peptide portion thereof, operably linked to a DNA binding domain of a transcription factor.

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Additional examples of invention isolated oligonucleotides, are those which generally are at least about 15 nucleotides in length and can hybridize specifically to the polynucleotide of the invention, but not to a polynucleotide encoding an N-CoR polypeptide (SEQ ID NO: 11). An oligonucleotide of the invention can be useful as a probe, or as a primer for a PCR procedure, or can encode a peptide containing at least five contiguous amino acids of a SMRT co-repressor. In one embodiment, an oligonucleotide of the invention encodes at least five contiguous amino acids of a sequence such as that shown as amino acids 720 to 745 of SEQ ID NO: 5; or amino acids 716 to 742 of SEQ ID NO: 7; or amino acids 497 to 523 of SEQ ID NO: 9. In another embodiment, an oligonucleotide of the invention can hybridize specifically to a polynucleotide encoding human SMRT (SEQ ID NO: 5) or mouse SMRTα (SEQ ID NO: 7), and, optionally, to a polynucleotide encoding mouse SMRTβ (SEQ ID NO: 9).

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The phrase "substantially the same" as used herein in reference to a nucleotide sequence of DNA, a ribonucleotide sequence of RNA, or an amino acid

sequence of protein, means sequences that have slight and non-consequential sequence variations from the actual sequences disclosed herein. Species that are substantially the same are considered to be equivalent to the disclosed sequences and as such are within the scope of the appended claims. In this regard, "slight and non-consequential sequence variations" means that sequences substantially the same as the DNA, RNA, or proteins disclosed and claimed herein are functionally equivalent to the sequences disclosed and claimed herein. Functionally equivalent sequences will function in substantially the same manner to produce substantially the same compositions as the nucleic acid and amino acid compositions disclosed and claimed herein. In particular, functionally equivalent DNAs encode proteins that are the same as those disclosed herein or that have conservative amino acid variations, such as substitution of a non-polar residue for another non-polar residue or a charged residue for a similarly charged residue. These changes include those recognized by those of skill in the art as those that do not substantially alter the tertiary structure of the protein.

In another embodiment according to the present invention, there are provided vectors comprising an invention polynucleotide, and host cells containing invention polynucleotides. The invention vector can be an expression vector, including, for example, a viral vector, and the polynucleotide, or a vector containing the polynucleotide, can be contained in a host cell. In one embodiment, the polynucleotide of the invention is operably linked to a tissue specific DNA regulatory element. In another embodiment, a SMRT co-repressor or peptide portion thereof encoded by the polynucleotide is expressed in a host cell.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates the repressor potential of a SMRT co-repressor. In this embodiment, the invention method comprises contacting a host cell with an agent, and detecting a change in the level of expression of a first expressible nucleotide sequence in response to the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor. In such a method, the host cell is characterized, in part, in that it contains a first expressible nucleotide

sequence operably linked to a first DNA regulatory element, and expresses a fusion polypeptide composed of an invention SMRT co-repressor, or peptide portion thereof, and a DNA binding domain of a first transcription factor that can specifically bind the first DNA regulatory element. Binding of the DNA binding domain of the first transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence in the host cell.

In another embodiment according to the present invention, there are provided methods for identifying an agent that modulates a function of an invention SMRT co-repressor. In this embodiment, the invention method comprises contacting an invention SMRT co-repressor, a member of the nuclear receptor superfamily, and an agent, and detecting an altered activity of the SMRT co-repressor in the presence of the agent as compared to the absence of the agent, thereby identifying an agent that modulates a function of the SMRT co-repressor.

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A method of the invention can be performed, for example, by contacting a host cell with an agent, and detecting a change in the level of expression of a first expressible nucleotide sequence in response to the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor. In such a method, the host cell is characterized, in part, in that it contains a first expressible nucleotide sequence operably linked to a first DNA regulatory element, and expresses a fusion polypeptide composed of a SMRT co-repressor or peptide portion thereof of the invention, and a DNA binding domain of a first transcription factor, which can specifically bind the first DNA regulatory element; binding of the DNA binding domain of the first transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence in the host cell. The first expressible nucleotide sequence can be an endogenous gene, which is normally present in the host cell, or can be a sequence that has been introduced into the host cell, either transiently or stably, using methods of recombinant DNA technology. In one embodiment, the first DNA binding domain is a GAL4 DNA binding domain and the first DNA regulatory element is a GAL4 DNA regulatory element that is operably

linked to an expressible nucleotide sequence, for example, a reporter gene, and is introduced into the host cell.

Thus, the invention method can identify an agent that increases or decreases the repressor potential of the SMRT co-repressor, or of an agent that increases or decreases the function of the SMRT co-repressor. The agent can directly interact with the SMRT co-repressor or peptide portion thereof, thereby modulating the repressor potential or function of the SMRT co-repressor, or can interact with a cellular molecule that, in turn, can alter the repressor potential or function of a SMRT co-repressor, thereby increasing or decreasing the repressor potential of the SMRT co-repressor.

The host cell can optionally contain a second expressible nucleotide sequence operably linked to a second DNA regulatory element, and can express a second fusion polypeptide, which is composed of an N-CoR polypeptide, or a repressor domain thereof, and a DNA binding domain of a second transcription factor, which can specifically bind the second DNA regulatory element. By comparing the level of expression of the first expressible nucleotide sequence and the second expressible nucleotide sequence in the host cell upon contacting the host cell with the agent, an agent that independently or coordinately modulates SMRT and N-CoR repressor activity. For example, detecting a change in the level of expression of the first expressible nucleotide sequence, but not in the level of expression of the second expressible nucleotide sequence, due to contacting the host cell with the agent identifies an agent that modulates the repressor potential of a SMRT co-repressor, but not of an N-CoR polypeptide can be identified.

In practicing a method of the invention, the SMRT co-repressor, or peptide portion thereof, can be, for example, an amino acid sequence such as amino acids 1 to 1031 of SEQ ID NO: 5; amino acids 1 to 1031 of SEQ ID NO: 7; or amino acids 1 to 813 of SEQ ID NO: 9. The agent can be, for example, an antibody or antigen binding fragment thereof, a peptide, or a small organic molecule.

In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing an invention isolated polynucleotide into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.

In another embodiment according to the present invention, there are provided methods of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing an invention isolated polynucleotide into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.

In performing a method of the invention, an agent that alters an interaction of the SMRT co-repressor, or peptide portion thereof, with the nuclear receptor can be identified using a binding assay, such as an electrophoretic mobility shift assay wherein the level of expression of an expressible nucleotide sequence. Such a method can also identify an agent that alters the ability of the invention SMRT co-repressor, or peptide portion thereof, to interact specifically with the nuclear receptor, but does not alter the level of expression of the expressible nucleotide sequence; or an agent that alters the level of expression of the expressible nucleotide sequence, but does not alter interaction of the SMRT co-repressor or peptide portion thereof with the nuclear receptor; or an agent that alters an interaction of the SMRT co-repressor, or peptide portion thereof, with the nuclear receptor and alters the level of expression of the expressible nucleotide sequence. The agent can, but need not be, a ligand for the nuclear receptor, and the method can be performed in a cell or in a reaction mixture *in vitro*.

Alternatively, an invention polynucleotide can be introduced into the cell, whereby the polynucleotide, or an expression product of the polynucleotide, alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor. The polynucleotide can encode an invention SMRT co-repressor or peptide, portion thereof, which can be expressed in the cell, thereby increasing the level of a SMRT co-repressor, or peptide portion thereof, in the cell. The polynucleotide also can be an antisense polynucleotide, that decreases the level of a SMRT co-repressor in the cell.

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In another embodiment according to the present invention, there are provided methods of identifying a molecule that interacts specifically with a SMRT co-repressor. In this embodiment, invention methods comprise contacting the molecule with an invention SMRT co-repressor and detecting specific binding of the molecule to the SMRT co-repressor, thereby identifying a molecule that interacts specifically with a SMRT co-repressor.

The molecule can be any molecule that interacts specifically with a SMRT co-repressor, including, for example, a small organic molecule such as a drug, a peptide, a nucleic acid molecule, and the like. In one embodiment, the molecule is a cellular factor, for example, a cellular protein that modulates the ability of a SMRT co-repressor to repress transcriptional activity of a nuclear receptor. In another embodiment, the method further involves isolating the molecule that interacts specifically with the SMRT co-repressor or peptide portion thereof.

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In accordance with yet another aspect of the present invention, there are provided methods to block the repressing effect of invention SMRT co-repressors, said method comprising administering an effective amount of an antibody as described herein. Alternatively, a silencing domain of a nuclear receptor can be employed. Those of skill in the art can readily determine suitable methods for administering said antibodies, and suitable quantities for administration, which will vary depending on

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numerous factors, such as the indication being treated, the condition of the subject, and the like.

In accordance with another aspect of the present invention, there is provided a method to repress (or silence) the activity of a member of the nuclear receptor superfamily containing a silencing domain that represses basal level promoter activity of target genes, said method comprising contacting said member of the nuclear receptor superfamily with a sufficient quantity of an invention SMRT co-repressor so as to repress the activity of said member. Members of the nuclear receptor superfamily contemplated for repression in accordance with this aspect of the present invention include, for example, thyroid hormone receptor, retinoic acid receptor, vitamin D receptor, peroxisome proliferator activated receptor, and the like.

In accordance with yet another aspect of the present invention, there is

provided a method to identify compounds which relieve the repression of nuclear
receptor activity caused by an invention SMRT co-repressor, said method comprising
comparing the size of the SMRT co-repressor/dimeric receptor complex (i.e., complexes
comprising the invention SMRT co-repressor and a homodimeric or heterodimeric
member of the nuclear receptor superfamily) upon exposure to test compound, relative to
the size of said complex in the absence of test compound. An observed size
corresponding to intact complex is indicative of an inactive compound, while an
observed size that reflects dissociation of the complex is indicative of a compound that
disrupts the complex, thereby relieving the repression caused thereby. Optionally, the
complex employed in this assay further comprises a response element for said member
of the nuclear receptor superfamily.

The size of the above-described complex can readily be determined employing various techniques available in the art. For example, electrophoretic mobility shift assays (EMSA) can be employed (wherein receptor alone or receptor-SMRT corepressor complex is bound to target DNA and the relative mobility thereof determined).

Those of skill in the art can readily identify other methodology which can be employed to determine the size of the complex as a result of exposure to putative ligand.

In accordance with a still further aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear receptor activity caused by an invention SMRT co-repressor, without substantially activating said receptor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer,

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter gene, and optionally, invention SMRT co-repressor, and

wherein said second expression system comprises a complex comprising:

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes,

the same response element-reporter combination as employed in said first expression system, and

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optionally, invention SMRT co-repressor, and thereafter selecting those compounds which provide:

a higher reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of said compound, and

substantially the same reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of an invention SMRT co-suppressor but substantially lacking the ability to activate nuclear receptor activity.

The addition of invention SMRT co-repressor is optional in the above-described assay because it is present endogenously in most host cells employed for such assays. It is preferred, to ensure the presence of a fairly constant amount of SMRT co-repressor, and to ensure that SMRT co-repressor is not a limiting reagent, that SMRT co-repressor be supplied exogenously to the above-described assays.

Mutant receptors contemplated for use in the practice of the present invention are conveniently produced by expression plasmids, introduced into the host cell by transfection. Mutant receptors contemplated for use herein include RAR403 homodimers, RAR403-containing heterodimers, TR160 homodimers, TR160-containing heterodimers, and the like.

Reporter constructs contemplated for use in the practice of the present invention comprise:

- (a) a promoter that is operable in the host cell,
- (b) a hormone response element, and

(c) a DNA segment encoding a reporter protein,
wherein the reporter protein-encoding DNA segment is
operatively linked to the promoter for transcription of the DNA
segment, and

wherein the hormone response element is operatively linked to the promoter for activation thereof.

Hormone response elements contemplated for use in the practice of the present invention are well known in the art, as has been noted previously.

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Exemplary reporter genes include chloramphenicol transferase (CAT), luciferase (LUC), beta-galactosidase ( $\beta$ -gal), and the like. Exemplary promoters include the simian virus (SV) promoter or modified form thereof (e.g., SV), the thymidine kinase (TK) promoter, the mammary tumor virus (MTV) promoter or modified form thereof (e.g.,  $\Delta$ MTV), and the like [see, for example, Mangelsdorf et al., in Nature 345:224-229 (1990), Mangelsdorf et al., in Cell 66:555-561 (1991), and Berger et al., in J. Steroid Biochem. Molec. Biol. 41:733-738 (1992).

As used herein in the phrase "operative response element" or

"operatively linked" the word "operative" means that the respective DNA sequences
(represented by the terms "GAL4 response element" and "reporter gene") are
operational, i.e., work for their intended purposes; such that after the two segments are
linked, upon appropriate activation by a ligand-receptor complex, the reporter gene will
be expressed as the result of the fact that the "GAL4 response element" was "turned on"
or otherwise activated.

In practicing the above-described functional bioassay, the expression plasmid and the reporter plasmid are co-transfected into suitable host cells. The transfected host cells are then cultured in the presence and absence of a test compound to determine if the test compound is able to produce activation of the promoter operatively linked to the response element of the reporter plasmid. Thereafter, the transfected and

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cultured host cells are monitored for induction (i.e., the presence) of the product of the reporter gene sequence.

Any cell line can be used as a suitable "host" for the functional bioassay contemplated for use in the practice of the present invention. Thus, cells contemplated for use in the practice of the present invention include transformed cells, non-transformed cells, neoplastic cells, primary cultures of different cell types, and the like. Exemplary cells which can be employed in the practice of the present invention include Schneider cells, CV-1 cells, HuTu80 cells, F9 cells, NTERA2 cells, NB4 cells, HL-60 cells, 293 cells, Hela cells, yeast cells, and the like. Preferred host cells for use in the functional bioassay system are COS cells and CV-1 cells. COS-1 (referred to as COS) cells are monkey kidney cells that express SV40 T antigen (Tag); while CV-1 cells do not express SV40 Tag. The presence of Tag in the COS-1 derivative lines allows the introduced expression plasmid to replicate and provides a relative increase in the amount of receptor produced during the assay period. CV-1 cells are presently preferred because they are particularly convenient for gene transfer studies and provide a sensitive and well-described host cell system.

The above-described cells (or fractions thereof) are maintained under physiological conditions when contacted with physiologically active compound. "Physiological conditions" are readily understood by those of skill in the art to comprise an isotonic, aqueous nutrient medium at a temperature of about 37°C.

In accordance with yet another aspect of the present invention, there is provided a method to identify compounds which activate nuclear receptor activity, but substantially lack the ability to relieve the repression caused by an invention SMRT corepressor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex

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comprising: a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor 5 homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer, a response element for said member of the nuclear receptor superfamily, wherein said response element is 10 operatively linked to a reporter, and optionally, invention SMRT co-repressor, and wherein said second expression system comprises a complex comprising: 15 a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, 20 the same response element-reporter combination as employed in said first expression system, and optionally, invention SMRT co-repressor, and thereafter selecting those compounds which provide: 25 a higher reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of compound, and substantially the same reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in

the absence of said compound,

wherein said selected compounds are capable of activating nuclear receptor activity, but substantially lacking the ability to relieve the repression caused by a SMRT co-repressor having a structure and function characteristic of, an invention SMRT co-repressor for retinoic acid and thyroid receptors.

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In accordance with a still further aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear receptor activity caused by an invention SMRT co-repressor, and activate said receptor, said method comprising:

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comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

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a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer,

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a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and optionally, invention SMRT co-repressor, and

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wherein said second expression system comprises a complex comprising:

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a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes,

# the same response element-reporter combination as employed in said first expression system, and optionally, invention SMRT co-repressor, and thereafter

5	selecting those compounds which provide:
	increased reporter signal upon exposure of said compound to said
	second expression system, relative to reporter signal in the absence of
	said compound, and
	substantially increased reporter signal upon exposure of said
10	compound to said first expression system, relative to reporter signal in
	the absence of said compound,
	wherein said selected compounds are capable of relieving the repression
	of nuclear receptor activity caused by a SMRT co-repressor having a structure and
15	function characteristic of the silencing mediator for retinoic acid and thyroid receptors,
	and activating said receptor.
	In accordance with still another embodiment of the present invention,
	there are provided modified forms of the above-described SMRT co-repressor,
20	including:
	full length silencing mediator for retinoic acid and thyroid receptors plus
	GAL4 DNA binding domain,
	full length silencing mediator for retinoic acid and thyroid receptors plus
	GAL4 activation domain,
25	full length silencing mediator for retinoic acid and thyroid receptors plus
	glutathione S-transferase (GST) tag,
	and the like.

The above-described modified forms of invention SMRT co-repressor can be used in a variety of ways, e.g., in the assays described herein.

An especially preferred modified SMRT co-repressor of the invention comprises full length silencing mediator for retinoic acid and thyroid receptors plus GAL4 activation domain.

5	In accordance with a still further embodiment of the present invention,
	there is provided a method to identify compounds which disrupt the ability of an
	invention SMRT co-repressor to complex with nuclear receptors, without substantially
	activating said receptor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above, a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer or retinoic acid receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

wherein said second expression system comprises a complex comprising:

said modified SMRT co-repressor,

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost

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its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

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selecting those compounds which provide:

a lower reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of said compound, and

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substantially the same reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of disrupting the ability of

a SMRT co-repressor having a structure and function characteristic of the silencing
mediator for retinoic acid and thyroid receptors to complex with nuclear receptors,
without substantially activating said receptor.

Mutant receptors contemplated for use in this embodiment of the present invention include RAR403 homodimers, RAR403-containing heterodimers, TR160 homodimers, TR160-containing heterodimers, and the like.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT (i.e., silencing mediator (SMRT corepressor) for retinoic acid receptor (RAR) and thyroid hormone receptor (TR)) is not endogenous to yeast.

In accordance with yet another embodiment of the present invention, there is provided a method to identify compounds which activate nuclear receptor activity, but substantially lack the ability to disrupt a complex comprising a nuclear receptor and an invention SMRT co-repressor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above, a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer or retinoic acid receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

wherein said second expression system comprises:

said modified SMRT co-repressor,

a homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

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selecting those compounds which provide:

a higher reporter signal upon exposure of said compound to said second expression system, relative to reporter signal in the absence of compound, and

substantially the same reporter signal upon exposure of said compound to said first expression system, relative to reporter signal in the absence of compound,

wherein said selected compounds are capable of activating nuclear receptor activity, but substantially lack the ability to disrupt the complex of an invention SMRT co-repressor.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT is not endogenous to yeast.

In accordance with a still further embodiment of the present invention, there is provided a method to identify compounds which activate a nuclear receptor, and disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:

comparing the reporter signal produced by two different expression systems in the absence and presence of test compound,

wherein said first expression system comprises a complex comprising:

a modified SMRT co-repressor as described above, a homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer or retinoic acid receptor-retinoid X receptor heterodimer, and

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a reporter, and

5 wherein said second expression system comprises a complex comprising:

said modified SMRT co-repressor,

the same homodimeric or heterodimeric member of the nuclear receptor superfamily as employed in said first expression system, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, and

the same response element-reporter combination as employed in said first expression system, and thereafter

selecting those compounds which provide:

a reduction in reporter signal upon exposure of compound to said first expression system, relative to reporter signal in the absence of said compound, and

increased reporter signal upon exposure of compound to said second expression system, relative to reporter signal in the absence of said compound,

wherein said selected compounds are capable of activating a nuclear receptor and disrupting a complex comprising nuclear receptor and a SMRT corepressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors.

Suitable host cells for use in this embodiment of the present invention include mammalian cells as well as yeast cells. Yeast cells are presently preferred because they introduce no background since SMRT is not endogenous to yeast.

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In accordance with yet another aspect of the present invention, there is provided a method to identify compounds which activate a nuclear receptor and/or disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:

comparing the reporter signals produced by a combination expression system in the absence and presence of test compound,

wherein said combination expression system comprises:

a first homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer,

a second homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first homodimer or heterodimer, wherein said member is mutated such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes (i.e., provides basal level expression),

> wherein either said first homodimer (or heterodimer) or said second homodimer (or heterodimer) is operatively linked to a GAL4 DNA binding domain,

a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a first reporter,

a GAL4 response element, wherein said response element is operatively linked to a second reporter, and

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optionally a SMRT co-repressor of nuclear receptor activity, said SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, and thereafter

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identifying as capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, but substantially lacking the ability to activate nuclear receptor activity those compounds which provide:

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a higher reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and

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substantially the same reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, or

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identifying as capable of activating nuclear receptor activity, but substantially lacking the ability to relieve the repression caused by a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors those compounds which provide:

> a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of compound, and

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substantially the same reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, or

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identifying as capable of relieving the repression of nuclear receptor activity caused by a SMRT co-repressor having a structure and function characteristic of

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the silencing mediator for retinoic acid and thyroid receptors, and activating said receptor those compounds which provide:

a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, and a greater increase in reporter signal from the reporter responsive

to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound.

Thus, the change in expression level of the two different reporters introduced in a single transfection can be monitored simultaneously. Based on the results of this single transfection, one can readily identify the mode of interaction of test compound with the receptor/SMRT complex.

Exemplary GAL4 response elements are those containing the palindromic 17-mer:

#### 5'-CGGAGGACTGTCCTCCG-3' (SEQ ID NO:3),

- such as, for example, 17MX, as described by Webster et al., in *Cell* **52**:169-178 (1988), as well as derivatives thereof. Additional examples of suitable response elements include those described by Hollenberg and Evans in *Cell* **55**:899-906 (1988); or Webster et al. in *Cell* **54**:199-207 (1988).
- In accordance with still another embodiment of the present invention, there is provided a method to identify compounds which activate a nuclear receptor and/or disrupt the ability of an invention SMRT co-repressor to complex with said receptor, said method comprising:
- comparing the reporter signals produced by a combination expression system in the absence and presence of test compound,

wherein said combination expression system comprises: a modified SMRT co-repressor as described above, a first homodimeric or heterodimeric member of the nuclear receptor superfamily selected from thyroid hormone receptor homodimer, thyroid hormone receptor-retinoid X 5 receptor heterodimer, retinoic acid receptor homodimer, or retinoic acid receptor-retinoid X receptor heterodimer, a second homodimeric or heterodimeric form of the same member of the nuclear receptor superfamily as employed in said first homodimer or heterodimer, wherein said member is mutated 10 such that it retains hormone dependent activation activity but has lost its ability to repress basal level promoter activity of target genes, wherein either said first homodimer (or heterodimer) or said second homodimer (or heterodimer) 15 is operatively linked to a GAL4 DNA binding domain, a response element for said member of the nuclear receptor superfamily, wherein said response element is operatively linked to a first reporter, 20 a GAL4 response element, wherein said response element is operatively linked to a second reporter, and thereafter identifying as capable of disrupting the ability of a SMRT co-repressor 25

identifying as capable of disrupting the ability of a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors to complex with a nuclear receptor, without substantially activating nuclear receptor, those compounds which provide:

a lower reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and substantially the same reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound, or

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identifying as capable of activating nuclear receptor activity, but substantially lacking the ability to disrupt a complex comprising a nuclear receptor and a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, those compounds which provide:

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a higher reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of compound, and

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substantially the same reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, or

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identifying as capable of disrupting a complex comprising a nuclear receptor and a SMRT co-repressor having a structure and function characteristic of the silencing mediator for retinoic acid and thyroid receptors, and activating said receptor those compounds which provide:

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a reduction in reporter signal from the reporter responsive to the first member upon exposure of said compound to said first member, relative to reporter signal in the absence of said compound, and

increased reporter signal from the reporter responsive to the second member upon exposure of said compound to said second member, relative to reporter signal in the absence of said compound.

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In accordance with a still further aspect of the present invention, there is provided a method to identify compounds which relieve the repression of nuclear

receptor activity caused by an invention SMRT co-repressor, said method comprising determining the effect of adding test compound to an expression system comprising:

a modified member of the nuclear receptor superfamily, wherein said
modified member contains an activation domain which renders said receptor
constitutively active,

a fusion protein comprising the receptor interaction domain of SMRT operatively linked to the GAL4 DNA binding domain, and

a GAL4 response element operatively linked to a reporter.

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Prior to addition of an effective ligand for the member of the nuclear receptor superfamily employed herein, the association of the modified member and the fusion protein will be effective to bind the GAL4 response element and activate transcription of the reporter. The presence of an effective ligand is indicated by a reduction of reporter signal upon exposure to ligand, which disrupts the interaction of the modified member and fusion protein.

Activation domains contemplated for use in the practice of the present invention are well known in the art and can readily be identified by the artisan. Examples include the GAL4 activation domain, BP64, and the like.

To summarize, a novel family of nuclear receptor SMRT co-repressor which mediates the transcriptional silencing of RAR and TR has been identified. This discovery is of great interest because transcriptional silencing has been shown to play an important role in development, cell differentiation and the oncogenic activity of v-erbA (Baniahmad et al., *EMBO J.* 11:1015-1023 (1992)); Gandrillon et al., *Cell* 49:687-697 (1989)); Zenke et al., *Cell* 61:1035-1049 (1990); Barlow et al., *EMBO J.* 13:4241-4250 (1994); Levine and Manley, *Cell* 59:405-408 (1989); Baniahmad et al., *Proc. Natl. Acad. Sci. USA* 89:10633-10637 (1992b); and Saitou et al., *Nature* 374:159-162 (1995)). In fact, v-erbA mutants that harbor the Pro160->Arg change in the TR neither repress basal

transcription nor are capable of oncogenic transformation (Damm and Evans, (1993), supra).

The function of SMRT as a silencing mediator (co-repressor) of RAR and TR is analogous to mSin3 in the Mad-Max-Sin3 ternary complex (Schreiber-Agus et al., Cell 80:777-786 (1995); and Ayer et al., Cell 80:767-776 (1995)). Because GAL-SMRT functions as a potent repressor when bound to DNA, it is reasonable to speculate that the function of the unliganded receptors is to bring with them SMRT to the template via protein-protein interaction. Thus, the repressor function is intrinsic to SMRT as opposed to the TR or RAR itself (Baniahmad et al., Proc. Natl. Acad. Sci. USA 10 90:8832-8836 (1993); and Fondell et al., Genes Dev 7:1400-1410 (1993)). It is demonstrated herein that the ligand triggers a dissociation of SMRT from the receptor, which would lead to an initial step in the activation process. This would be followed (or be coincident) with an induced conformational change in the carboxy-terminal transactivation domain ( c, also called AF2), allowing association with co-activators 15 on the transcription machinery (Douarin et al., EMBO J. 14:2020-2033 (1995); Halachmi et al., Science 264:1455-1458 (1994); Lee et al., Nature 374:91-94 (1995); and Cavailles et al., Proc. Natl. Acad. Sci. USA 91:10009-10013 (1994)). Thus, as has previously been suggested (Damm and Evans, (1993), supra), the ligand dependent activation of TR would represent two separable processes including relief of repression 20 and net activation. The isolation of SMRT now provides a basis for dissecting the molecular basis of trans-repression.

The invention will now be described in greater detail by reference to the following non-limiting examples.

#### Example 1 Isolation of SMRT

Using a GAL4 DBD-RXR fusion protein (see, for example, USSN 30 08/177,740, incorporated by reference herein in its entirety) as a bait in a yeast

two-hybrid screening system (Durfee et al., (1993), *supra*), several cDNA clones encoding receptor interacting proteins were isolated. One of these proteins, SMRT, interacts strongly with unliganded RAR and TR but only weakly with RXR or other receptors in yeast. This protein was selected for further characterization.

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### Example 2 Far-western blotting procedure

Total bacteria extracts expressing GST fusions of hRARα (aa 156-462)

or hRXRα LBD (aa 228-462) and control extracts expressing GST alone or GST-PML fusion protein were subjected to SDS/PAGE and electroblotted onto nitrocellulose in transfer buffer (25 mM Tris, pH 8.3/ 192 mM glycine/ 0.01% SDS). After denaturation/renaturation from 6 M to 0.187 M guanidine hydrochloride in HB buffer (25 mM HEPES, pH 7.7/25 mM NaCl/5 mM MgCl<sub>2</sub>/1 mM DTT) filters were saturated at 4°C in blocking buffer (5% milk, then 1% milk in HB buffer plus 0.05% NP40). *In vitro* translated <sup>35</sup>S-labeled proteins were diluted into H buffer (20 mM Hepes, pH 7.7/75 mM KCl/0.1 mM EDTA/2.5 mM MgCl<sub>2</sub>/0.05% NP40/ 1% milk/1 mM DTT) and the filters were hybridized overnight at 4°C with (1 μM) or without ligand. After three washes with H buffer, filters were dried and exposed for autoradiography or quantitated by phosphoimager.

GST-SMRT is a GST fusion of the C-SMRT encoded by the yeast two hybrid clone. GST-SMRT has been purified, but contains several degradation products.

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For yeast two-hybrid screening, a construct expressing the GAL4 DBD-hRXRα LBD (aa 198-462) fusion protein was used to screen a human lymphocyte cDNA library as described (Durfee et al., (1993), *supra*). Full length SMRT cDNA was isolated from a human HeLa cDNA library (Clontech) using the two-hybrid insert as a probe.

Using the above-described far-western blotting procedure, <sup>35</sup>S-labeled SMRT preferentially complexes with bacterial extracts expressing the RAR, marginally associates with RXR and shows no association with control extracts. In contrast, <sup>35</sup>S-PPAR selectively associates with its heterodimeric partner, RXR, but not with RAR. In a similar assay, <sup>35</sup>S-labeled RAR or TR interacts strongly with SMRT and their heterodimeric partner, RXR, but not with degraded GST products, while <sup>35</sup>S-RXR interacts only weakly with SMRT. Binding of ligand to RAR or TR reduces their interactions with SMRT but not with RXR, while binding of ligand to RXR has only slight effect. Figure 1 shows the quantitation of a dose-dependent dissociation of SMRT from RAR or TR by all-*trans* retinoic acid (atRA) or thyroid hormone (triiodothyronine or T3), demonstrating that the amount of ligand required for 50% dissociation in both cases are close to the kds for both ligands (Munoz et al. *EMBO J.* 7:155-159 (1988); Sap et al., *Nature* 340:242-244 (1989); and Yang et al., *Proc. Natl. Acad. Sci. USA* 88:3559-3563 (1991)).

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Full length SMRT encodes a polypeptide of 1495 amino acids rich in proline and serine residues (see Figure 2 and SEQ ID NO:1). Genbank database comparison reveals similarity of the C-terminal domain of SMRT to a partial cDNA encoding another receptor interacting protein, RIP13 (Seol et al., (1995), *supra*), whose role in receptor signaling is unknown. Within this region, there can be identified several potential heptad repeats which might mediate protein-protein interaction with the "a-helical sandwich" structure (Bourguet et al., *Nature* 375:377-382 (1995)) of the ligand binding domain (LBD) of receptors.

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### Example 3 Characterization of SMRT

Unlike other nuclear receptors, unliganded RAR and TR possess a strong silencing domain which represses basal level promoter activity of their target genes (Damm et al., *Nature* **339**:593-597 (1989); Brent et al., *New Biol.* **1**:329-336 (1989); Baniahmad et al., *Cell* **61**:505-514 (1990); and Baniahmad et al., *EMBO J.* 

11:1015-1023 (1992)). The preferential interaction of SMRT with RAR and TR in the absence of hormone suggests that SMRT may play a role in mediating the transcriptional silencing effect of the receptor.

To further investigate the involvement of SMRT in silencing, the interaction of SMRT with mutant receptors which display distinct silencing and/or transactivation activities was tested as follows. <sup>35</sup>S-methionine labeled receptors were used as probes to hybridize immobilized GST-SMRT in the presence (10 μM) or absence of all-*trans* retinoic acid (atRA). The total bacteria extract expressing

10 GST-RXR was included as a control.

When quantitated by phosphoimager, RAR403 shows a 4-fold better interaction with SMRT than wild type RAR. Both full length RAR or a deletion mutant expressing only the ligand binding domain (LBD, referred to as  $\Delta\Delta$ R) associate with SMRT; this association is blocked by ligand.

These results confirm that the LBD alone is sufficient in the interaction. The carboxy-terminal deletion mutant RAR403 is a potent dominant negative repressor of basal level promoter activity of RAR target genes (Damm et al., *Proc. Natl. Acad. Sci. USA* 90:2989-2993 (1993); Tsai and Collins, *Proc. Natl. Acad. Sci. USA* 90:7153-7157 (1993); and Tsai et al., *Genes Dev* 6:2258-2269 (1992)). As might be predicted from the above studies, RAR403 and its amino terminal deletion derivative, R403, interact strongly with SMRT in either the presence or absence of ligand, consistent with SMRT mediating the repressor activity of this mutant.

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## Example 4 Interaction of SMRT with TR Mutants

The interaction of SMRT with two different classes of TR mutants was analyzed next. The first mutant employed is the naturally occurring oncogene, v-erbA, which has strong silencing ability but no transactivation activity (Sap et al., (1989),

supra; Sap et al., Nature 324:635-640 (1986); Weinberger et al., Nature 318:670-672 (1985); and Weinberger et al., Nature 324:641-646 (1986)). The second mutant employed is a single amino acid change (Pro 160 -> Arg) of the rTRa (TR160) which has previously been shown to lose its capacity in basal level repression but retains hormone dependent transactivation (Thompson et al., Science 237:1610-1614 (1987); and Damm and Evans, Proc. Natl. Acad. Sci. USA 90:10668-10672 (1993)). If SMRT is involved in silencing, it would be expected that SMRT should interact with the v-erbA, but show little or no association with the silencing-defective TR160 mutant.

Interaction of the oncogenic v-erbA and rTR $\alpha$  R160 mutant (TR160) with GST-SMRT was determined in a far-western assay as described above (see Example 2). When quantitated by phosphoimager, the v-erbA shows an 18-fold better interaction with SMRT than hTR $\beta$ , and the TR160 mutant shows a 10-fold lower signal than the rTR $\alpha$ .

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As one might expect, v-erbA interacts strongly with SMRT both in presence or absence of ligand. In contrast, full length TR160 mutant or LBD of TR160 ( $\Delta\Delta$ TR160) does not interact significantly with SMRT when compared to the wild type receptor.

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These data demonstrate that SMRT plays an important role in mediating transcriptional silencing effects of both RAR and TR. These data also suggest that the release of SMRT from receptors could be a prerequisite step in ligand-dependent transactivation by nuclear receptors.

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## Example 5 Formation of ternary complexes containing SMRT

RAR and TR form heterodimers with RXR, resulting in a complex with 30 high DNA binding ability (Bugge et al., *EMBO J.* 11:1409-1418 (1992); Yu et al., *Cell* 67:1251-1266 (1991); and Kliewer et al., *Nature* 355:446-449 (1992)). Since SMRT

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interacts with RAR and TR, tests were conducted to determine whether SMRT can also interact with the receptor-DNA complex. Thus, the interaction of SMRT with RXR-RAR heterodimer on a DR5 element (i.e., an AGGTCA direct repeat spaced by five nucleotides) was determined in a gel retardation assay, which is carried out as follows. *In vitro* translated receptor or unprogrammed reticulocyte lysate (URL) was incubated with 1 µg of poly dIdC on ice for 15 minutes in a total volume of 20 µl containing 75 mM KCl, 7.5% glycerol, 20 mM Hepes (pH 7.5), 2 mM DTT and 0.1% NP-40, with or without ligand (in the range of about 10-100 nM employed). A <sup>32</sup>P labeled, double stranded oligonucleotide probe was added into the binding reaction (10,000 cpm per reaction), and the reaction was further incubated for 20 minutes at room temperature. The protein-DNA complex was separated on a 5% native polyacrylamide gel at 150 volts.

SMRT is seen to form a ternary complex with the RXR-RAR heterodimer on a DNA response element in the gel retardation assay. Addition of ligand releases SMRT from this complex in a dose-dependent manner.

Similarly, SMRT is seen to form a ternary complex with the RXR-TR heterodimer on a TR response element; addition of T3 disrupts the formation of this complex.

These data demonstrate that SMRT can be recruited to DNA response elements via protein-protein interaction with RAR or TR in the absence of hormone. Binding of hormone disrupts receptor-SMRT interaction and releases SMRT from the receptor-DNA complex.

#### Example 6

#### Transient transfection assay

30 CV-1 cells were plated in 24 well plates at a density of 50,000 cells per well. Expression plasmids were transfected into cells by lipofection using DOTAP. In

each transfection, 5 ng of GAL-RAR and 15 ng of v-erbA or SMRT were used together with 150 ng of reporter construct containing 4 copies of GAL4 binding sites in front of a minimal thymidine kinase promoter and a CMX- $\beta$ -gal construct as an internal control. The relative luciferase activity was calculated by normalizing to the  $\beta$ -gal activity.

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### Example 7 Reversal of transcriptional silencing

Recently, it has been shown that over expression of RAR or TR could reverse the transcriptional silencing effect of the GAL4 DBD fusion of TR (GAL-TR) or RAR (GAL-RAR) (Baniahmad et al., *Mol Cell Biol* 15:76-86 (1995); and Casanova et al., *Mol Cell Biol* 14:5756-5765 (1994)), presumably by competition for a limiting amount of a SMRT co-repressor. A similar effect is observed herein when over expression of v-erbA or RAR403 mutants are shown to reverse the silencing effect of GAL-RAR and GAL-TR on the basal activity of a luciferase reporter (see Figure 3A and 3B).

In principle, over expression of SMRT should restore repressor activity when co-expressed with v-erbA or RAR403 competitors. Indeed, results presented in Figure 3C show that both the full length and the C-terminal domain of SMRT (C-SMRT) can titrate out v-erbA or RAR403 competitor activity and re-endow GAL-RAR and GAL-TR with silencing activity. In contrast, neither v-erbA nor SMRT show any effect on the transactivation activity of GAL-VP16 fusion. Thus, SMRT is able to block the titration effect of v-erbA and RAR403 and functionally replaces the putative SMRT co-repressor in this system.

## Example 8 Direct recruitment of SMRT to a heterologous promoter

30 If SMRT is the mediator of transcription silencing of TR and RAR by interaction with template-bound unliganded receptors, then direct recruitment of SMRT

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to a heterologous promoter should result in repression of basal level activity. This was tested by fusing full length SMRT to the GAL4 DBD (GAL-SMRT). The effect of the resulting fusion protein on the activity of the thymidine kinase promoter containing four GAL4 binding sites was analyzed. Figure 3D shows that GAL-SMRT, like GAL-TR, can silence basal promoter activity in a dose-dependent manner. In contrast, GAL-RXR shows no repression.

These data suggest that SMRT, when recruited to a promoter by direct DNA binding or via association with an unliganded receptor, functions as a potent transcriptional repressor.

#### Example 9

#### Cloning Of Human And Mouse SMRT co-repressors

This example describes the cloning of a full length human silencing mediator of retinoic acid and thyroid hormone receptor (SMRT co-repressor) and of two mouse SMRT isoforms, m-SMRT $\alpha$  and m-SMRT $\beta$ .

An examination of the previously described human SMRT co-repressor revealed that the first eight amino acids and upstream sequences were derived from a portion of ribonucleoprotein K sequence. Accordingly, a mouse spleen cDNA lambda ZAP II library (Stratagene; La Jolla CA) was screened at low stringency with a probe corresponding to approximately the 5' 1,000 base pairs (bp) of the previously identified human SMRT (s-SMRT). A 3.5 kilobase (kb) cDNA fragment was obtained that contained a unique sequence in addition to known s-SMRT sequence. The 5' end of this cDNA, and subsequently obtained clones, was used in successive rounds of screening of the mouse spleen cDNA library and a mouse brain cDNA library (Stratagene) and the full-length SMRTα isoform cDNA (SEQ ID NO: 6) and SMRTα isoform cDNA (SEQ ID NO: 10) were obtained. The mouse SMRT (m-SMRT) 5' sequence then was used at low stringency to screen a human pituitary cDNA library (Stratagene) to obtain the full-length human SMRT (h-SMRT) cDNA (SEQ ID NO: 1). All cDNA clones were

sequenced on both strands using standard methods, and have been deposited with GenBank as Accession No. AF103003 (h-SMRT; SEQ ID NOS: 3 and 5); Accession No. 113001 (m-SMRTα; SEQ ID NOS: 6 and 7); and Accession No. 113002 (m-SMRTβ; SEQ ID NOS: 8 and 9).

By sequentially shifting between the mouse spleen and mouse brain cDNA libraries, several clones containing a potential starting methionine and 5' untranslated region sequences were obtained. The complete polypeptide sequences of m-SMRT (SEQ ID NO: 7) and h-SMRT (SEQ ID NO: 5) are provided. In addition, a splice variant isolated from the mouse brain cDNA library encoded an m-SMRT corepressor containing a deletion of amino acids 36 to 254 of SEQ ID NO: 7 (see SEQ ID NO: 3). The two m-SMRT co-repressors are designated SMRT $\alpha$  (SEQ ID NO: 7) and SMRT $\beta$  (SEQ ID NO: 9). Based on sequence similarity to N-CoR (see below), this deletion in m-SMRT $\beta$  removes the majority of the sequence in h-SMRT and m-SMRT $\alpha$  that is homologous to N-CoR repression domain 1 (RD1), including a portion of the Sin3A binding region.

The cloned h-SMRT (SEQ ID NO: 3) encodes a polypeptide that contains an additional 1130 amino acids at the amino terminus as compared to the previously described human SMRT co-repressor. The full length h-SMRT shares 84% identity with m-SMRTα. A comparison of h-SMRT (SEQ ID NO: 5) and N-CoR (SEQ ID NO: 11) revealed that the N-terminal extension of h-SMRT (amino acids 1 to 1030) and N-CoR (amino acids 1 to 1031) share approximately 41% identity, which is somewhat higher that the 36% identity shared between the full length proteins. However, regions within the N-CoR and SMRT N-termini share striking homology (Figures 4A and 4B).

Amino acids 1 to 160 of N-CoR are moderately conserved in h-SMRT (and m-SMRTα), sharing about 36% identity. This region of N-CoR has been reported to interact with Siah2 (Zhang et al., (1998), *supra*) and, similarly, can be involved in an

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interaction of Siah2 with h-SMRT or m-SMRT $\alpha$ . In particular, highly conserved sequences in this region can be the specific Siah2 interaction sites (see Figure 4A).

A 52 amino acid segment from N-CoR (amino acids 255 to 312) mediates an interaction with Sin3A (Heinzel et al., *Nature* **387**:43-48 (1997)), and was presumed to represent the core of the larger RD1 region (Horlein et al., (1995), *supra*). This small interaction domain is highly conserved (about 83% identity) in h-SMRT, and the overall identity shared between SMRT and N-CoR RD1 is about 57%.

Amino acids 312 to 668 of N-CoR also are well conserved (66% identity) in h-SMRT (and m-SMRTα), and two internal blocks of sequences in this region share even greater similarity (see Figure 1B; shaded regions). These blocks are homologous to each other and to part of the SANT domain, which was identified in the yeast chromatin remodeling factor, SWI3, the yeast adapter protein, ADA2, the basal
transcription factor TFIIIB, and other proteins (Aasland et al., *Trends Biochem. Sci.*21:87-88 (1996)), suggesting that these domains share a common and important function. The amino acids of N-CoR RD2 (see Horlein et al., (1995) *supra*) are the least conserved in h-SMRT, sharing about 30% identity.

These results demonstrate that isoforms of SMRT co-repressors are expressed in cells, as exemplified by m-SMRT $\alpha$  and m-SMRT $\beta$ . In addition, the results demonstrate that the previously undescribed amino terminus of SMRT co-repressors shares regions of substantial homology with N-CoR, and regions of homology are identified that indicate these sequences can mediate previously uncharacterized functions.

#### Example 10

#### Expression And Chromosomal Localization Of Smrt Co-Repressors

This example describes the tissue distribution of SMRT RNA and the chromosomal localization of human SMRT.

Total RNA was prepared from adult CB6F1 mouse tissues using TRIZOL reagent (GIBCO/BRL), and poly(A) RNA was purified from total RNA using an OLIGOTEX mRNA Kit (Qiagen, Valencia, CA). RNA was separated on 1.25% agarose/6% formaldehyde gels and transferred to a NYTRAN membrane (Scheicher & Schuell). A 720 bp m-SMRT/PstI fragment was used as a probe. Following hybridization with the SMRT probe, the filters were stripped and hybridized with a murine glyceraldehyde-3-phosphate dehydrogenase cDNA probe to allow normalization for RNA loading.

Chromosomal localization of SMRT was determined by fluorescence in situ hybridization using the 5.3 kb h-SMRT cDNA clone. The probe was labeled by nick-translation with biotin-11-dUTP, then hybridized to normal male human metaphase chromosomes. Chromosomes were counterstained with 4',6-diamidino-2-phenylindole (DAPI). Chromosome identification was carried out by computer inversion of the gray scale DAPI image on a PSI Imaging System (Perceptive Scientific Instruments; League City TX). Chromosome 12 confirmation was carried out using a chromosome 12-specific alpha satellite probe (Vysis; Downers Grove IL).

Previous studies using the short human SMRT co-repressor suggested that SMRT was expressed ubiquitously in various tissues. To confirm this result, expression of the full length m-SMRT was determined by northern blot analysis by using a probe consisting of nucleotides 2760 to 3620 of m-SMRT (SEQ ID NO: 6). The expression pattern was ubiquitous, as previously described, although higher levels were detected in lung, spleen, and brain. Similarly, h-SMRT was expressed ubiquitously as determined using a multiple tissue blot (CLONTECH; Palo Alto CA). It is noteworthy that two isoforms of SMRT were present in the majority of the mouse tissues and likely correspond to the m-SMRTα and m-SMRTβ isoforms.

The chromosomal location of the h-SMRT and N-CoR genes was mapped. The h-SMRT clone hybridized to the q arm of one of the C group

chromosomes. Computer-mediated banding of the DAPI stained chromosomes identified the labeled chromosome as chromosome 12, band q24. The chromosome 12 localization was confirmed by cohybridization of SMRT and a chromosome 12 alpha satellite probe, D12Z3 (Vysis), which labels the pericentromeric region of chromosome 12. The location for the human N-CoR gene was determined through a mapped human bacterial artificial chromosome clone, hCIT529I10, which is 158 kb of genomic N-CoR and resides on chromosome 11p11.2. The SMRT and N-CoR chromosomal locations can be accessed through GENEMAP98 from the Human Genome Project at http://www.ncbi.nlm.nih.gov/genemap.

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These results demonstrate that the full length SMRT co-repressors and the SMRT co-repressors are expressed in various tissues. The results also demonstrate that the human SMRT gene is located on chromosome 12.

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# Example 11 Functional Characterization Of SMRT Amino Terminus Domains

This example demonstrates that various domains of the SMRT amino terminus can repress nuclear receptor transcriptional activity.

Experiments were performed using the plasmids pCMX-GAL4 DBD and pMH100-TK-luc (Nagy et al., (1997), *supra*). Standard PCR amplifications were used to generate GAL4 fusion constructs. All constructs were verified by double-stranded sequencing to confirm identity and reading frame.

Monkey CV-1 cells were grown in DMEM supplemented with 10% resin-charcoal stripped fetal bovine serum (FBS), 50 units/ml of penicillin G, and 50  $\mu$ g/ml of streptomycin sulfate at 37°C in 7% CO<sub>2</sub>. V-1 cells (60-70% confluence, 48-well plate) were cotransfected with 16 ng of pCMX-GAL4, 100 ng of pMH100-TK-luc, and 100 ng of pCMX- $\beta$  galactosidase in 200  $\mu$ l of DMEM containing 10% super-

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stripped fetal calf serum (FCS) by the N-(1-(2,3-dioleoyloxy)propyl)-N,N,N-trimethylammonium methylsulfate (DOTAP)-mediated procedure (Nagy et al., (1997), supra). The amount of DNA in each transfection was kept constant by addition of pCMX. After 24 hr, the medium was replaced; cells were harvested and assayed for luciferase activity 36 to 48 hr after transfection. Luciferase activity was normalized by the level of  $\beta$ -galactosidase activity. Each transfection was performed in triplicate and repeated at least three times.

Based on the high degree of identity between regions of the SMRT amino terminus and the corresponding N-CoR region, the ability of regions in the SMRT amino terminus to act in transcriptional repression was examined. A nested series of nucleotide sequences encoding portions of the SMRT amino terminus fused to the GAL4 DNA binding domain (GAL-DBD) was prepared in mammalian expression vectors (Figure 5A). The constructs were cotransfected with a GAL4-TK-luciferase reporter plasmid to determine the regulatory properties of the GAL4-SMRT fusions. Repression was determined relative to the basal activity of the reporter in the presence of the GAL-DBD alone.

The entire SMRT amino terminus region (GAL4-SMRT(1-1031))

demonstrated the greatest amount of repression (approximately 38-fold), and virtually extinguished reporter activity. In comparison, GAL4-SMRT (1-303), which is equivalent to N-CoR RD1, demonstrated 6-fold repression; and GAL4-SMRT (736-1031), which is equivalent to N-CoR RD2, demonstrated about 2.6-fold repression. Surprisingly, the highly conserved SANT domain conferred a significant amount of repression (about 3.3-fold).

A smaller region (amino acids 845 to 986) within the RD2 homology region shows a higher level of sequence conservation as compared to the entire RD2 region. Deletion constructs were generated to determine whether this minimal region was sufficient for the repression activity of RD2. Deletion of flanking amino acids 736 to 845 or of amino acids 987 to 1055 did not affect the level of repression, demonstrating

that the repressor function of RD2 is contained within a 141 amino acid core sequence of RD2.

Based on sequence similarity to N-CoR, the deletion of amino acids 36 to 254 in the m-SMRT $\beta$  isoform removes the majority of RD1, including a portion of the Sin3A binding region. The effect of this deletion on SMRT function was examined by cotransfection experiments comparing repression by SMRT $\alpha$  to SMRT $\beta$ . These experiments revealed that SMRT $\beta$  has substantially less repressor activity than SMRT $\alpha$ . A construct containing the entire amino terminus of m-SMRT $\beta$  (amino acids 1 to 813) repressed activity about 2.6 fold, as compared to m-SMRT $\alpha$  amino acids 1 to 1031, which repressed activity about 38.1-fold. In addition, a GAL4 construct containing m-SMRT amino acids 1 to 83 repressed activity only about 1.4-fold. These results indicate that alternative splicing can add further diversity to expand the function of SMRT gene products.

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### Example 12 Yeast Two-Hybrid Screen and Assays

To investigate whether repression by EcR in CV-1 cells is mediated by its association with a vertebrate corepressor and whether such an interaction, if it does occur, is impaired by the A483T mutation, a mammalian two-hybrid assay with Gal4-c-SMRT was conducted.

A yeast two-hybrid screen (Fields and Song, *Nature*, **340**:245-246, (1989)) was performed by transforming approximately 2 X 10<sup>6</sup> Y190 yeast cells with a pAS-EcR construct and a Drosophila (0-8 hr) embryonic c-DNA two-hybrid library (Yu et al., *Nature*, **385**:552-555, (1997)). Transformants were selected onto DO-Leu-Trp-His plates containing 40 mM 3-aminotriazole (Sigma) for 3-4 days. Surviving yeast colonies were picked as primary positives and restreaked on selection plates to isolate single clones. Activation domain plasmids were rescued from the selected positive transformants for further analysis. Each clone was evaluated by testing its

potential interaction with several other nuclear receptors using the yeast two-hybrid assays. E52 was isolated and further pursued based on this selection criterion. Quantitative liquid assay of  $\beta$ -galactosidase was performed on positive clones 16 hr after treating the yeast cells with no ligand, or with 3  $\mu$ M ligand.

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pAS-EcR is a fusion gene with the region corresponding to amino acids 223-878 of EcRB1 fused C-terminally to the Gal4-DBD of the pAS1-CYH2 construct (Durfee et al., (1993), *supra*); other Gal4-DBD-based nuclear receptor constructs used in this yeast two-hybrid assay include: USP (amino acids 50-508), hRAR (amino acids 186-462) and hTR (amino acids 121-410) (Schulman et al., (1995), *supra*), and SMRT (Chen and Evans, (1995), *supra*). β-galactosidase activities were quantified by liquid assay for yeast cells treated either without ligand or with 3 μM of corresponding hormone. All-trans retinoic acid (ATRA) is a ligand of RAR; 3,3',5-triiodothyroacetic acid (T3) is a ligand of TR.

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Similar yeast two-hybrid assays were also used to examine the interaction between SMRTER and mSin3A and dSin3A.

### Example 12 Cloning SMRTER

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To isolate full-length SMRTER cDNA, a XhoI insert fragment isolated from the E52 clone was used to screen male and female Tudor c-DNA libraries (gift of Tulle Hazelrigg). This initial screen resulted in isolating three overlapping c-DNA clones covering the region of amino acid 2094 to the C terminus of SMRTER Additional regions were obtained from three consecutive library screens using two cosmid clones isolated from the Tamkun genomic library (gift of John Tamkun). Sequences of these overlapping c-DNA and genomic clones were assembled to obtain a conceptual open reading frame of SMRTER 3446 amino acids in length (SEQ ID NO:12; Figure 8A). The translational initiation codon was designated based on the sequences that match the consensus Kozak codons and is preceded by three in-frame

consecutive stop codons in the upstream region. Both strands of the sequences of the c-DNA clones were determined using an ABI prism Big Dye® terminator cycle sequencing ready reaction kit (PE Biosystems) and ABI 377 instrument.

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### Example 14 Plasmids

CMV promoter-driven expression plasmids of EcR, USP, RXR,

10 c-SMRT, β-galactosidase, and pMH100-TK-luc reporter, and yeast plasmids of RAR,

TR, and SMRT have been described previously (Yao et al., (1992), *supra*, Yao et al.,

(1993), *supra*; Chen and Evans, (1995), *supra*; Schulman et al., (1995), *supra*; Chen

et al., *Proc. Natl. Acad. Sci. USA* 93:7567-7571, (1996); Nagy et al., (1997), *supra*).

hsp27EcR-TK-Luc, a reporter with six copies of the hsp27EcRE, is a gift of Barry

15 Forman. CMV vector-driven EcR A483T and Gal4-SMRD3 mutations were

generated using the Transformer® site-directed mutagenesis kit (Clontech) with

proper selection primers and the mutagenic primers that correspond to the missense

mutation (A483T) of EcR and to the designated mutations, M2 and M3, in the

SMRD3 domain, respectively. Other plasmids were constructed with standard

20 techniques, including various enzyme digestions or PCR amplification.

### Example 15 Cell Culture and Transfection

25 CV-1 cells were grown in Dulbecco's modified Eagles medium at 37°C in 5% CO<sub>2</sub>. The media were supplemented with 10% AG1-X8 resin charcoal double-stripped calf bovine serum, 50 U/ml penicillin G, and 50 µg/ml streptomycin sulfate. Approximately 20 hr after CV-1 cells (10<sup>5</sup> cells) were plated in 48-well cell culture clusters (Costar), cells were transiently transfected with plasmids using DOTAP according to the instructions of the manufacturer (Boehringer Mannheim). The amount of CMV promoter-driven expression vectors, β-galactosidase gene

expression vector, CMX-lacZ, and reporter, pMH100-TK-luc or hsp27EcRE-TK-Luc, were in the range of 100-200 ng, 500 ng, and 400 ng, respectively, for six wells of each 48-well clusters in each transfection experiments. At least 4 hr after transfection, each medium was replaced with medium either without ligand, or with 1 µM of MurA. Cells were harvested and assayed approximately 48 hr after transfection. All experiments were performed in triplicate and repeated with similar results.

CV-1 cells were transfected with wild-type EcR or EcR A483T, along with vp16-USP and a reporter, hsp27EcRE-TK-Luc, which contains six copies of the hsp27EcRE fused to the thymidine kinase (TK) promoter-luciferase reporter. VP16-USP fusion contains the region of USP (amino acids 50-508) fused C-terminally to the VP16 domain. Muristerone A (MurA) is a potent ecdysone agonist (Christopherson et al., *Proc. Natl. Acad. Sci. USA,* 89:6314-6318, (1992)). In all experiments, cells were also cotransfected with CMV-lacZ, which is used to normalize the luciferase activity. As shown in Figure 6A, the ability to dimerize with USP is reflected in reporter activity without treatment with hormone (open bar), and the ability to respond to hormone is reflected in reporter activity when cells were treated with 1 μM Muristerone A (closed bar).

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CMV promoter-driven expression vector including wild-type EcR or EcR A483T was cotransfected with VP16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), *supra*) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine—threonine) at the 483 site of EcR (Bender et al., (1997), *supra*). The results of this experiment (Figure 6B) show that EcR A483T disrupts the interaction with SMRT.

### Example 16 In Vitro Interacting Assays

Glutathione S-transferase fusion proteins, including GST-X, GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1), and GST-ERID2 (amino acids 2951-3038 of SEQ ID NO:1), were expressed in E. coli DH5 cells, and extracts were affinity purified by binding to glutathione Sepharose 4B beads. Bound proteins used as affinity matrices in pull-down experiments were first equilibrated with the binding buffer (20 mM HEPES [pH 7.9], 150 mM NaCl, 1 mM EDTA, 4 mM MgCl2, 1 mM DTT, 0.06% NP40, 10% Glycerol, 0.25 mM PMSF, 1 mg BSA). For pull-down assays using GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) and GST-ERID2 (amino acids 2951-3038 of SEQ ID NO:1), additional hsp27EcRE (0.05 µg/ml) was added to the binding buffer. In this experiment, 30 µl of 50% GST-protein beads slurry, containing approximately 1 µg of proteins, were incubated with 10 µl of 35Smethionine-labeled proteins in 300  $\mu l$  of the binding buffer (with or without 3  $\mu M$  of MurA as indicated) for 30 min at room temperature. After the incubation, beads were washed three times with the binding buffer (with or without ligand) and resuspended in SDS-PAGE sample buffer before loading. After electrophoresis, bound radiolabeled proteins were visualized by autoradiography. 35S-methionine-labeled EcR, USP were generated in a coupled transcription-translation system, TNT (Promega), using CMX-EcR (T7) and CMX-uspK (T7) constructs as templates, respectively.

# Example 17 Immunohistochemistry and Immunofluorescence

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Antibodies against SMRTER were raised in rabbits immunized with bacterially expressed glutathione S-transferase fusion proteins corresponding to the region (amino acids 2477-2648 of SEQ ID NO:1) of SMRTER. Specific antibodies were purified by affinity chromatography through antigen-linked columns and used at 1:200 dilution for tissue staining. Tissues for whole-mount staining were dissected at the wandering third instar stage of the Canton-S strain larvae and fixed (4%

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formaldehyde in 1? PBS, 50 mM EGTA) for at least 30 min. Preincubation, secondary antibodies, washes, and peroxidase reactions are described in the protocol of the Elite ABC (Rabbit IgG) kit (Vector). For the pilot experiments, partially purified IgG from preimmunization serum was used. For polytene chromosome staining, salivary glands were dissected according to the method described in Zink et al., *EMBO J.*, **10**:153-162, (1991).

Chromosome spreads were costained with affinity-purified anti-SMRTER (1:100) polyclonal antibody and with anti-USP monoclonal antibody (ABIII/AD5; gift of F. Kafatos, 1:100 dilution). SMRTER was detected with Texas red-conjugated donkey anti-rabbit secondary antibody (1:100 dilution), and USP was detected with FITC-conjugated donkey anti-mouse secondary antibody (1:100 dilution) (Jackson ImmunoResearch Labs).

### 15 <u>Example 18</u> ER Interacts Genetically with DSinA

In keeping with the evidence that dSin3A is a component in EcR regulatory pathway, an experiment was conducted to examine whether dSin3A interacts genetically with EcR using several previously characterized Drosophila EcR and dSin3A mutants (Bender et al., (1997), *supra*; Neufeld et al., (1998), *supra*). In the experiment, in which female dSin3AK07401 were crossed with male EcRE261st using techniques known in the art (see Table 1 below), only approximately 14% of the scored EcRE261st/dSin3AK07401 progenies survived, a percent that is significantly lower than the expected 33.3%. This suggests that a large portion of the EcRE261st/dSin3AK07401 flies either die prior to eclosion or fail to eclose. Additionally, surviving EcRE261st/dSin3AK07401 escapers showed delayed development and wing defects, in which wings are held horizontally at 45°-90° angle from the body axis. These results suggest that dSin3A shares an overlapping regulatory pathway with EcR.

In a reverse genetic cross, in which female EcRE261st were crossed with male dSin3AK07401, none of the EcRE261st/dSin3AK07401 flies survived to adulthood. These results suggest that EcRE261st/dSin3AK07401 results in a genetically sensitized background. When the maternally deposited EcR in embryos descended from female EcRE261st/SM6b was cut in half, the lethality for EcRE261st/dSin3AK07401 was further increased. These results reveal that, in addition to its previously known zygotic function, EcR also contributes maternally to Drosophila development.

10 <u>Table 1</u>

Table 1. EcR Interacts Genetically with DSin3A		
		EcR <sup>E261st</sup> /DSin3A <sup>KO7401</sup>
Cross		Surviving Rate (%)
DSin3A <sup>KO7401</sup> /CyO	φ	
×		14 (n = 141)
EcR <sup>261st</sup> /SM6b	8	
EcR <sup>261st</sup> /SM6b	9	
×		0 (n = 144)
DSin3A <sup>KO7401</sup> /CyO	3	P261at va27/

A similar wing held-out phenotype is also observed in EcR<sup>E261st</sup>/DSin3A<sup>xe374</sup>, Df(2R)nap11/DSin3A<sup>KO7401</sup>, and Df(2R)nap11/Dsin2A<sup>xe374</sup>. EcR<sup>E261st</sup> and Df(2R)nap11 are both described in Figure 6. Dsin2A<sup>KO7401</sup> is an allele with a P element insertion within the 5' intron of Sin3A. DSin3A<sup>xe374</sup> is an X ray-generated allele (Neufeld et al., (1998)). n=the number of surviving flies scored. Note that CyO/SM6b is lethal.

EcRA483T flies showed developmental abnormalities in wings and tergites.

A similar phenotype, although with a lower penetration rate, has been also observed in EcRA483T/Df(2R)20B and in EcRA483T/Df(2R)nap11. Df(2)20B and Df(2)nap11 are both deficiencies in which EcR is deleted (Bender et al., (1997), supra). Sequence alignment of EcR with the vertebrate TR, RAR, and v-erbA, an oncogenic TR variant, revealed that alanine 483 is located within a highly conserved 23-amino acid (aa) loop region connecting helices 3 and 4, termed the LBD signature

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motif (Wurtz et al., *Nat. Struct. Biol.*, 3:206, (1996)) (see Figure 6C). Based on structural studies of vertebrate nuclear receptors (for review, see Moras and Gronemeyer, (1998), *supra*), this alanine residue appears to be on the exposed surface, consistent with it being a potential corepressor binding site for nuclear receptors.

These in vivo studies indicate that EcRA483T is a semilethal allele (Bender et al., (1997), *supra*). When EcRA483T is in trans with EcRE261st, an allele that removes both the DBD and LBD domains of EcR, animals are primarily lethal (>95%). The few surviving EcRA483T/EcRE261st flies, however, display significant delays in development, blistered wings, and defective tergites, indicating that EcR is involved in the development of these tissues. The ability of EcR to bind a vertebrate corepressor and the loss of this property in EcR A483T suggests that the defects observed in EcRA483T flies may result from the disruption of its interaction with an as yet unidentified Drosophila corepressor.

# Example 19 Isolation of an EcR-Interacting Factor

The CMV promoter-driven expression vector including wild-type EcR or EcR A483T, was cotransfected with vp16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), *supra*) into CV-1 cells to examine its effect on the interaction of the invertebrate SMRTER with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine→threonine) at the 483 site of EcR (Bender et al., (1997), *supra*). Although EcR readily interacted with SMRT in both mammalian and yeast cells (Figure 6B; Figure 7), repeated low-stringency hybridization screens failed to identify a Drosophila homolog of SMRT. No

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#### Example 20

### <u>Isolation and Characterization of an</u> EcR-Interacting Clone - Yeast Two-hybrid screen

To pursue the isolation of an EcR corepressor, a yeast two hybrid interaction screen was performed of a Drosophila embryonic cDNA library using pAS-EcR as bait. E52 was isolated as one of the complementary positive clones from a yeast two-hybrid screen with pAS-EcR as bait, as described in Example 12.

#### 10 <u>Example 21</u>

#### Characterization of a Repression-Defective EcR Allele, EcRA483T

(A) CV-1 cells were transfected with wild-type EcR or EcR A483T, along with vp16-USP and a reporter, hsp27EcRE-TK-Luc, which contains six copies of the hsp27EcRE fused to the thymidine kinase (TK) promoter-luciferase reporter. In all experiments, cells were also cotransfected with CMV-lacZ, which is used to normalize the luciferase activity. The ability to dimerize with USP was reflected in reporter activity without treatment with hormone (open bar), and the ability to respond to hormone was reflected in reporter activity when cells were treated with 1 μM Muristerone A (closed bar). vp16-USP fusion contains the region of USP (amino acids 50-508) fused C-terminally to the vp16 domain. Muristerone A (MurA) is a potent ecdysone agonist (Christopherson et al., (1992), *supra*). In these tests EcR A483T was selectively defective in repression.

or EcR A483T was cotransfected with vp16-USP and Gal4-c-SMRT (amino acids 981 to C terminus) (Chen and Evans, (1995), *supra*) into CV-1 cells to examine its effect on the interaction with vertebrate corepressor. All cells were also cotransfected with a TK-luciferase reporter construct, pMH100-TK-Luc, containing four copies of the yeast Gal4-responsive element. EcR A483T corresponds to a single amino acid change (alanine threonine) at the 483 site of EcR (Bender et al., (1997), *supra*). The

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results of this test show that EcR A483T disrupts the interaction with SMRT.

(C) Sequence alignment of EcR with the vertebrate TR, RAR, and verbA, an oncogenic TR variant, reveals that the alanine 483 of the EcRA4831T mutant is located within a highly conserved 23-amino acid (aa) loop region connecting helices 3 and 4, termed the LBD signature motif (Wurtz et al., (1996), supra) (Figure 6C). Based on structural studies of vertebrate nuclear receptors (for review, see Moras and Gronemeyer, (1998), supra), this alanine residue appears to be on the exposed surface, consistent with it being a potential corepressor binding site for nuclear receptors.

In vivo studies indicated that EcRA483T is a semilethal allele (Bender et al., (1997), *supra*). When EcRA483T is in trans with EcRE261st, an allele that removes both the DBD and LBD domains of EcR, animals are primarily lethal (>95%). The few surviving EcRA483T/EcRE261st flies, however, display significant delays in development, blistered wings, and defective tergites, indicating that EcR is involved in the development of these tissues. The ability of EcR to bind a vertebrate corepressor and the loss of this property in EcR A483T suggested to us that the defects observed in EcRA483T flies may result from the disruption of its interaction with an as yet unidentified Drosophila corepressor.

## Example 22 Isolation of an EcR-Interacting Factor

Although EcR readily interacts with SMRT in both mammalian and yeast cells (Figure 6B; Figure 7), repeated low-stringency hybridization screens failed to identify a Drosophila homolog of SMRT. Given that no SMRT/N-CoR homolog is found in C. elegans, it was believed that either a SMRT/N-CoR-like corepressor is not conserved in invertebrates or, alternatively, invertebrate corepressors may diverge significantly from their vertebrate counterparts. To pursue the isolation of an EcR corepressor, a yeast interaction screen of a Drosophila embryonic cDNA library using

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EcR as bait was conducted as described in Example 19. This screen resulted in the isolation of a clone, E52, whose protein product interacts with EcR as well as with the vertebrate RAR and TR, but notably not with USP (Figure 7). Unlike the interaction between E52 and RAR, which can be dissociated by all-trans retinoic acid, the interaction between E52 and EcR, or the interaction between SMRT and EcR, is not dissociated by Muristerone A (MurA). This result suggests that other factors essential for the dissociation of E52 from EcR, such as USP, are missing in yeast (see below).

#### Example 23

#### Isolation and Characterization of an EcR-Interacting Clone

E52 was isolated as one of the complementary positive clones from a yeast two-hybrid screen. Isolation of overlapping cDNA and genomic clones led to the identification of a full-length sequence encoding a large protein of 3446 amino acids (Figure 8A). This protein contains several unusually long stretches of Gln, Ala, Gly, and Ser repeats. Comparative analysis reveals it to be a novel protein with limited regions of clear homology with the vertebrate nuclear receptor corepressors SMRT and N-CoR (Chen and Evans, (1995), *supra*; Hörlein et al., (1995), *supra*; Ordentlich et al., (1999), *supra*; Park et al., (1999), *supra*). This protein SMRTER, SMRT-related ecdysone receptor-interacting factor, was shown by Northern blot analysis to encode large transcripts (>12 kb) expressed broadly throughout the embryonic stage and three larvae stages, as well as in adult Drosophila flies.

#### Example 24

#### Molecular and Biochemical Analysis for ERID1 and ERID2

Interaction with the EcR complex was evaluated based on transient transfection with the Gal4-SMRTER fusion genes. USP, EcR-vp16 (VP16 transactivating domain was fused C-terminally to the end of the EcRB1 isoform), and the reporter, pMH100-TK-Luc.

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In vitro pull down assays (Example 12) were conducted to determine whether EcR interacts with ERID1 and ERID2. In vitro translated 35S-methionine-labeled EcRB1 alone, or a mixture of 35S-methionine-labeled EcRB1 and unlabeled USP, or 35S-methionine-labeled USP alone, were incubated with GST, GST-ERID1 (amino acids 1698-2063 of SEQ ID NO:1), or GST-ERID2 (amino acids of SEQ ID NO:1). GST-ERID1 and GST-ERID2, but not GST alone, pull down labeled EcR, whereas little interaction is found between USP and any of the three GST proteins. In addition, the pull-down complex was disrupted by the addition of 3μM MurA when USP is present. These in vitro results establish that SMRTER and EcR may interact directly.

Further in vitro tests were conducted to determine ERID1, ERID2, and c-SMRT compete with each other to bind EcR. Gal4-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) or Gal4-ERID2 (amino acids 2929-3181 of SEQ ID NO:1), along with EcR-vp16 and USP, were transfected in CV-1 cells as described above. In this competition experiment, additional ERID1, ERID2, and c-SMRT (Chen et al., (1996), supra) were cotransfected into cells. ERID1 (1698-2063) and ERID2 ((amino acids 2929-3038 of SEQ ID NO:1) were tagged with the nuclear targeting signal (MAPKKKRKV) (SEQ ID NO:3) to ensure that these proteins were localized in nuclei. As shown in Figure 11C, interaction between each Gal4-ERID fusion and EcR-vp16:USP was significantly decreased by both ERIDs and by c-SMRT. Interestingly, a more prominent effect was observed in experiments when Gal4-ERID1 (amino acids 1698-2063 of SEQ ID NO:1) was challenged by ERID2, and, conversely, a more efficient competition was achieved by ERID1 to Gal4-ERID2 (amino acids 2094-3181 of SEQ ID NO:1). Together, these results suggest that ERID1, ERID2, and c-SMRT may bind similar or overlapping surface(s) in EcR.

## Example 25 SMRTER Colocalizes with the EcR on Polytene Chromosomes

SMRTER antibodies were prepared as described in Example 12 to examine its cytological and chromosomal localization patterns of SMRTER.

Consistent with its action as a corepressor of EcR, SMRTER was localized to nuclei of salivary glands and of fat bodies, as well as to nuclei of eye, wing, and leg imaginal discs isolated from the third instar larvae.

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Next association of SMRTER with the EcR:USP complex on chromosomes was examined. The USP staining pattern was used as an index for EcRs presence on chromosomes. Since USP and EcR colocalized with each other on polytene chromosomes (Yao et al., (1993), *supra*), chromosomal spreads prepared from the salivary glands of wandering third instar larvae (prior to pupariation) were subjected to simultaneous immunological staining with antibodies against SMRTER and USP. SMRTER was detected with antibody conjugated with Texas red, USP with FITC.

To visualize the band, interband, and puffing patterns of the polytene chromosomes, the chromosomes were counterstained with DAPI to show the banding regions while leaving the interbands and puffs unstained or lightly stained. Indirect immunofluorescence staining revealed that SMRTER is a chromosome-bound protein and colocalizes with USP (FITC) at a majority of chromosomal sites; whereas in a pilot experiment, no such staining patterns were detected using the preimmunization serum. The strongest SMRTER staining was primarily associated with the boundary between band and interband regions as well as within the interband regions of chromosomes counterstained with DAPI. This result confirms that, as an EcR-associating factor, SMRTER is recruited by the EcR:USP heterodimers to their specific target chromosomal loci.

SMRTER staining can still be detected in puffed regions, such as the 2B puff. Since the polytene chromosomes consist of a parallel arrangement of several hundred to two thousand copies of the euchromatic portions of the chromosomes, an individual binding protein like SMRTER may be cycling on and off, resulting in a steady state of signals detected in the broader chromatin regions. Whether or not SMRTER levels actually change prior to or after the peak of ecdysone pulses remains to be established.

While the invention has been described in detail with reference to certain preferred embodiments thereof, it will be understood that modifications and variations are within the spirit and scope of that which is described and claimed.

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#### That which is claimed is:

- 1. An isolated polynucleotide encoding a member of a family of silencing mediators of retinoic acid receptor and thyroid hormone receptor, or an isoform or peptide portion thereof (SMRT co-repressor), or an isolated polynucleotide complementary thereto.
- 2. The polynucleotide of claim 1, which modulates transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor).
- 3. The polynucleotide of claim 2, wherein the SMRT co-repressor comprises a repression domain having
  - a) less than about 83% identity with a Sin3A interaction domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
  - b) less than about 57% identity with repression domain 1 of N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11;
  - c) less than about 66% identity with a SANT domain of N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
  - d) less than about 30% identity with repression domain 2 of N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11, and polynucleotides that hybridize thereto under stringent conditions.
- 4. The polynucleotide of claim 1, wherein the SMRT co-repressor is a human SMRT co-repressor having an amino acid sequence as set forth in SEQ ID NO: 5 or conservative variations thereof.
- 5. A polynucleotide which hybridizes under stringent conditions with a polynucleotide according to claim 2.

- 6. A polynucleotide that has at least 80% sequence identity with a polynucleotide according to claim 2.
- 7. The polynucleotide of claim 4, which has a nucleotide sequence as set forth in SEQ ID NO: 4, and conservative variations thereof.
- 8. The polynucleotide of claim 1, wherein the SMRT co-repressor is a mouse SMRT $\alpha$  isoform.
- 9. The polynucleotide of claim 6, having an amino acid sequence as set forth in SEQ ID NO: 7 or conservative variations thereof.
- 10. The polynucleotide of claim 4, which has a nucleotide sequence as set forth in SEQ ID NO: 6.
- 11. The polynucleotide of claim 1, wherein the SMRT co-repressor is a mouse SMRT $\beta$  isoform.
- 12. The polynucleotide of claim 11, having an amino acid sequence as set forth in SEQ ID NO: 9 or conservative variations thereof.
- 13. The polynucleotide of claim 11, which has a nucleotide sequence as set forth in SEQ ID NO: 8.

14. The polynucleotide of claim 1, comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 3094 of SEQ ID NO: 4; nucleotides 1 to 3718 of SEQ ID NO: 6; and nucleotides 1 to 2801 of SEQ ID NO: 8.

15. A polynucleotide that under stringent conditions with a polynucleotide according to claim 14, provided that the polynucleotide does not contain a sequence identical to SEQ ID NO: 11.

- 16. A polynucleotide that has at least 80% sequence identity with a polynucleotide according to claim 14, provided that the polynucleotide does not contain a sequence identical to SEQ ID NO: 11.
- 17. A polynucleotide of claim 1, comprising a nucleotide sequence selected from the group consisting of:

nucleotides 1 to 8388 of SEQ ID NO: 6; and nucleotides 1 to 7465 of SEQ ID NO: 8.

- 18. The polynucleotide of claim 1, comprising nucleotides 1 to 8561 of SEQ ID NO: 4.
- 19. The polynucleotide of claim 1, which is operably linked to a second nucleotide sequence.

- 20. The polynucleotide of claim 19, which encodes a fusion polypeptide comprising the SMRT co-repressor operably linked to a DNA binding domain of a transcription factor.
  - 21. A vector comprising the polynucleotide of claim 1.
  - 22. A host cell containing the polynucleotide of claim 1.
- 23. An isolated oligonucleotide, comprising at least 15 nucleotides that can hybridize specifically to the polynucleotide of claim 1, but not to a polynucleotide encoding SEQ ID NO: 11 or to a polynucleotide encoding an amino acid sequence consisting of amino acids 1031 to 2517 of SEQ ID NO: 5.
- 24. The oligonucleotide of claim 23, wherein the polynucleotide encodes at least five contiguous amino acids of a sequence selected from the group consisting of:

amino acids 720 to 745 of SEQ ID NO: 5; amino acids 716 to 742 of SEQ ID NO: 7; and amino acids 497 to 523 of SEQ ID NO: 9.

25. The oligonucleotide of claim 23, which can hybridize specifically to a polynucleotide encoding SEQ ID NO: 5 or SEQ ID NO: 7, but not to a polynucleotide encoding SEQ ID NO: 9.

hormone receptor, or isoform or peptide portion thereof (SMRT co-repressor),

26. An isolated silencing mediator of retinoic acid and thyroid

	wherein the co-repressor modulates transcriptional potential of a member of the
	nuclear receptor superfamily (nuclear receptor).
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	27. An isolated co-repressor comprising a repression domain having
	a) less than about 83% identity with a Sin3A interaction
	domain of N-CoR set forth as amino acids 255 to 312 of SEQ ID NO: 11;
	b) less than about 57% identity with repression domain 1 of
10	N-CoR set forth as amino acids 1 to 312 of SEQ ID NO: 11;
	c) less than about 66% identity with a SANT domain of
	N-CoR set forth as amino acids 312 to 668 of SEQ ID NO: 11; or
	d) less than about 30% identity with repression domain 2 of
	N-CoR set forth as amino acids 736 to 1031 of SEQ ID NO: 11.
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	28. An isolated peptide, comprising at least six contiguous amino
	acids of an amino acid sequence selected from the group consisting of:
	amino acids 1 to 1030 of SEQ ID NO: 5;
	amino acids 1 to 1029 of SEQ ID NO: 7;
20	amino acids 1 to 809 of SEQ ID NO: 9;
	and conservative variations thereof,
	provided the peptide is not identical to a sequence of SEQ ID NO: 11.
	29. An isolated antibody that binds specifically to the peptide of claim
	28.

30. A cell line, which produces the antibody of claim 29.

26 and at least a second molecule.

31. A chimeric molecule, comprising the SMRT co-repressor of claim

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- 32. A complex, comprising a SMRT co-repressor of claim 26 and a member of the nuclear receptor superfamily (nuclear receptor).
- 33. The complex of claim 32, wherein the nuclear receptor is in the form of a dimer.
- 34. A method for identifying an agent that modulates the repressor potential of a SMRT co-repressor, the method comprising:
  - a) contacting a host cell with an agent,
    wherein the host cell contains a first expressible nucleotide
    sequence operably linked to a first DNA regulatory element, and
    expresses a fusion polypeptide comprising a SMRT corepressor of claim 26, and a DNA binding domain of a first transcription
    factor, which can specifically bind the first DNA regulatory element,

and wherein binding of the DNA binding domain of the first transcription factor to the first DNA regulatory element results in expression of the first expressible nucleotide sequence; and

- b) detecting a change in the level of expression of the first expressible nucleotide sequence due to contacting the host cell with the agent, thereby identifying an agent that modulates the repressor potential of a SMRT co-repressor.
- 35. A method for identifying an agent that modulates a function of a SMRT co-repressor, the method comprising:
  - a) contacting a SMRT co-repressor of claim 26, a member of the nuclear receptor superfamily (nuclear receptor), and

an agent; and

b) detecting an altered activity of the SMRT co-repressor in the presence of the agent as compared to the absence of the agent, thereby identifying an agent that modulates a function of the SMRT co-repressor.

- 36. A method of modulating the transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor) in a cell, the method comprising introducing a polynucleotide of claim 1 into the cell, whereby the polynucleotide or an expression product of the polynucleotide alters the level of a SMRT co-repressor in the cell, thereby modulating the transcriptional potential of the nuclear receptor.
- 37. A method of identifying a molecule that interacts specifically with a SMRT co-repressor, the method comprising:
- a) contacting the molecule with the SMRT co-repressor of claim 26; and
  - b) detecting specific binding of the molecule to the SMRT corepressor, thereby identifying a molecule that interacts specifically with a SMRT co-repressor.

## ABSTRACT OF THE INVENTION

The present invention relates to isolated polynucleotides encoding a family of silencing mediators of retinoic acid and thyroid hormone receptor (SMRT) isoforms, including vertebrate and invertebrate isoforms thereof. For example, a full length human SMRT co-repressor, two isoforms of a mouse SMRT-- a longer form, mouse SMRT $\alpha$  , and a shorter form, mouse SMRT $\beta$ , and an isoform of an insect (Drosophilia), SMRTER -- as well as peptide portions of the SMRT co-repressors that can modulate transcriptional potential of a member of the nuclear receptor superfamily (nuclear receptor); to oligonucleotides that can hybridize specifically to such a polynucleotide; to vectors and to host cells containing such polynucleotides. The invention also relates to polypeptide SMRT co-repressors encoded by such invention SMRT polynucleotides, and to peptide portions thereof that can modulate transcriptional potential of a nuclear receptor; including peptide portions of a SMRT co-repressor that are not present in an N-CoR polypeptide. In addition, the invention relates to chimeric molecules and to complexes containing a SMRT co-repressor or peptide portion thereof, to antibodies that specifically bind such compositions, and to methods for identifying an agent that modulates the repressor potential of a SMRT corepressor. The invention also provides methods for identifying an agent that modulates a function of a SMRT co-repressor; for modulating the transcriptional potential of a nuclear receptor in a cell using the compositions of the invention; and for identifying a molecule that interacts specifically with a SMRT co-repressor.

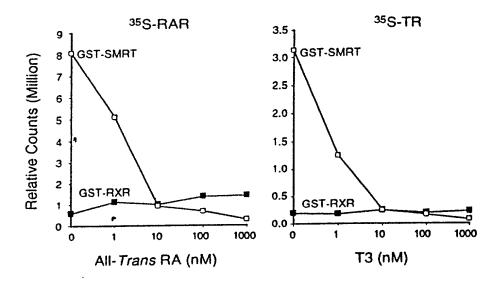


FIGURE 1

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401	RSIHEIPREELRHTPELPLAPRPLKEGSITQGTPLKYDTGASTTGSKKHD
451	VRSLIGSPGRTFPPVHPLDVMADARALERACYEESLKSRPGTASSSGGSI
501	ARGAPVIVPELGKPRQSPLTYEDHGAPFAGHLPRGSPVTMREPTPRLQEG
551	SLSSSKASQDRKLTSTPREIAKSPHSTVPEHHPHPISPYEHLLRGVSGVD
601	LYRSHIPLAFDPTSIPRGIPLDAAAAYYLPRHLAPNPTYPHLYPPYLIRG
651	YPDTAALENRQTIINDYITSQQMHHNTATAMAQRADMLRGLSPRESSLAL
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751	SSSPLSPGGPTHLTKPTTTSSS <u>ERERDRDRERDRDREREK</u> SILTSTTTVE
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1001	REKTQSKPFSIQELELRSLGYHGSSYSPEGVEPVSPVSSPSLTHDKGLPK
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	`
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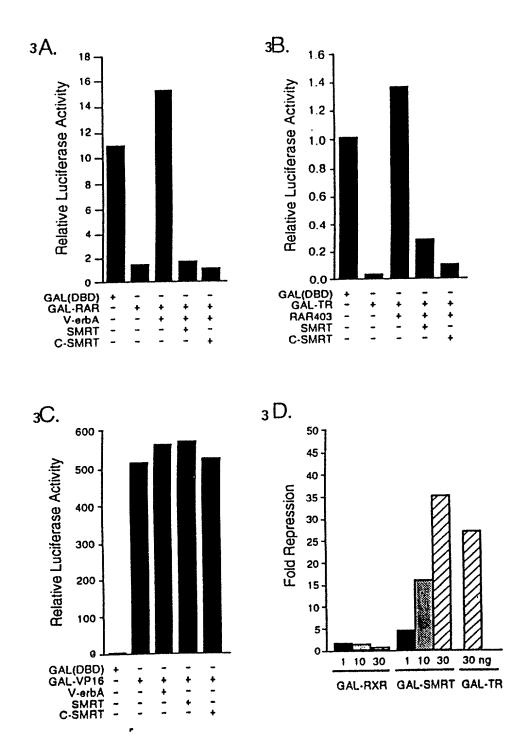


FIGURE 3

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# FIGURE 4

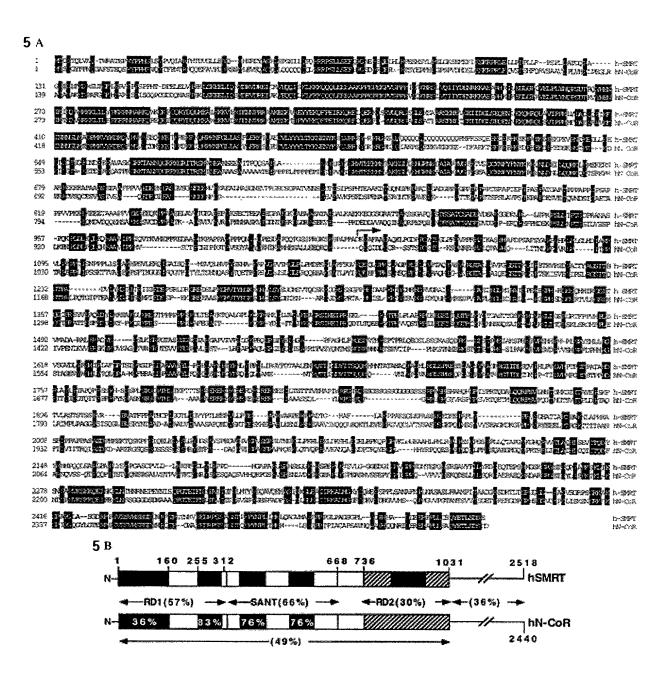
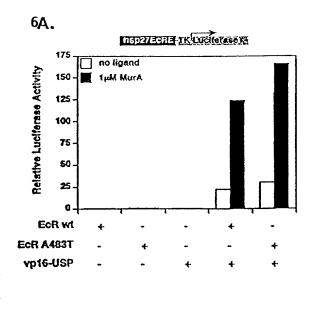
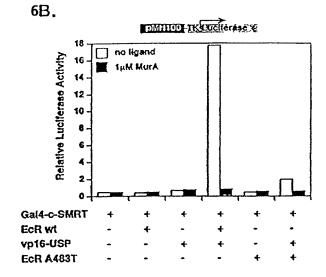
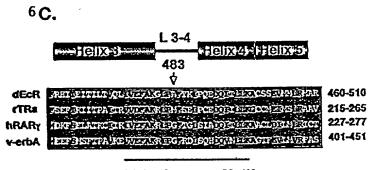


FIGURE 5







LBD-signature Motif

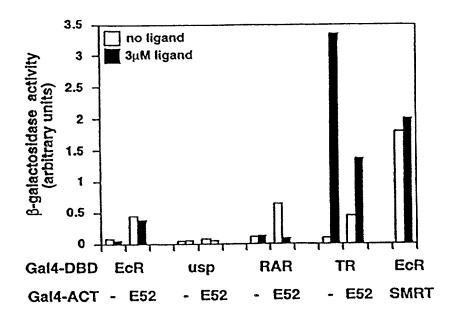


FIGURE 7

### **DECLARATION FOR PATENT APPLICATION**

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship is as stated below next to my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **A FAMILY OF TRANSCRIPTIONAL CO-REPRESSORS THAT INTERACT WITH NUCLEAR HORMONE RECEPTORS AND USES THEREFOR**, which is a C-I-P of 08/522,726, filed on September 1, 1995, the specification of which

X	is attached hereto. (SALK1510-3)	
<u>X</u>	was filed on March 10, 2000, as U.S. Application Serial No.	
	, and was amended on,	if
applic	able (the "Application").	

I hereby authorize and request insertion of the application serial number of the Application when officially known.

I hereby state that I have reviewed and understand the contents of the aboveidentified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, Code of Federal Regulations ("C.F.R."), § 1.56.

With respect to the Application, I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

(Application Serial No.)	(Filing Date)
(Application Serial No.)	(Filing Date)
(Application Serial No.)	(Filing Date)

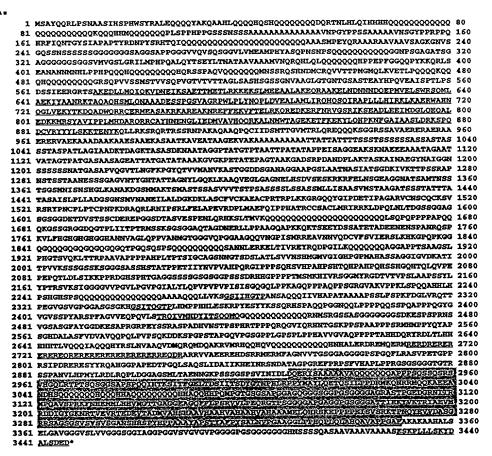
With respect to the Application, I hereby claim the benefit under 35 U.S.C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of the application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability of the subject matter of the Application as defined in Title 37, C.F.R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of the Application:

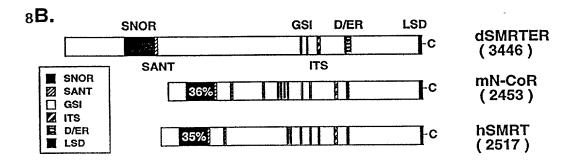
08/522,726 (Application Serial No.)		
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)
(Application Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so

made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of first inventor: Ronald M. Evans
Inventor's signature:
Date:
Residence: <u>La Jolla, California</u>
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Post Office Address: 1471 Cottontail Lane  La Jolla, California 92037
Full name of second inventor:
Inventor's signature:
Date:
Residence: San Diego, California
Citizenship: Taiwan
Post Office Address: 7548 Charmant Drive, #1416 San Diego, California 92126
Full name of third inventor: Peter Ordentlich
Inventor's signature:
Date:
Residence:
Citizenship:
Post Office Address:





# SNOR motif COURTE: 513 ALCORA 117 ALCO

FIGURE 9

LSD motif

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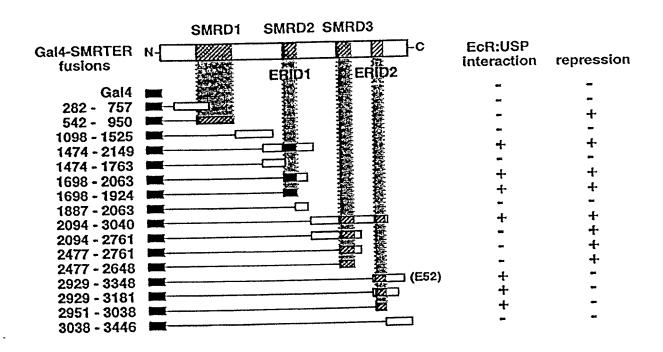
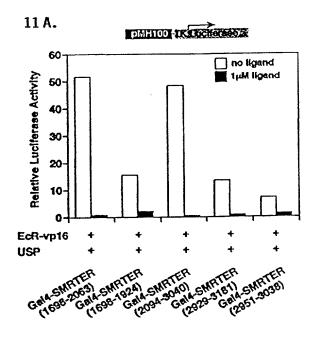


FIGURE 10



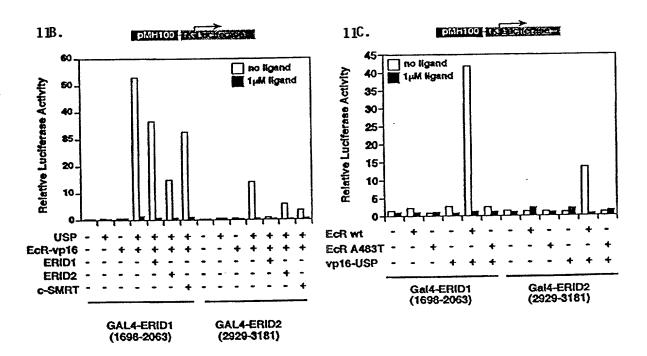
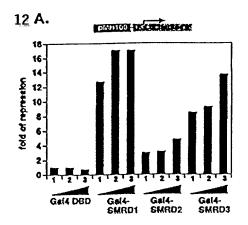
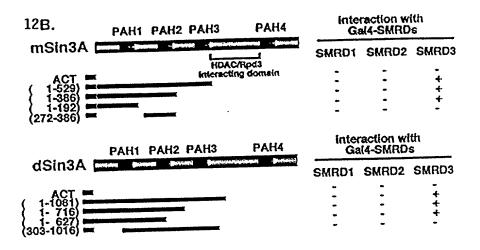
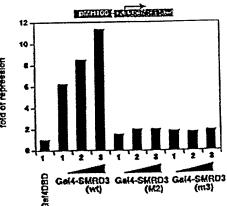


FIGURE 11





12 C.



### SEQUENCE LISTING

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acg cac acg gac gtc ggg ctc ctg gag tac cag cac cac tcc cgc gac Thr His Thr Asp Val Gly Leu Leu Glu Tyr Gln His His Ser Arg Asp 35 40 45	145													
tat gcc tcc cac ctg tcg ccg ggc tcc atc atc cag ccc cag cgg cgg Tyr Ala Ser His Leu Ser Pro Gly Ser Ile Ile Gln Pro Gln Arg Arg 50 55 60	193													
agg ccc tcc ctg ctg tct gag ttc cag ccc ggg aat gaa cgg tcc cag Arg Pro Ser Leu Leu Ser Glu Phe Gln Pro Gly Asn Glu Arg Ser Gln 65 70 75 80	241													
gag ctc cac ctg cgg cca gag tcc cac tca tac ctg ccc gag ctg ggg Glu Leu His Leu Arg Pro Glu Ser His Ser Tyr Leu Pro Glu Leu Gly 85 90 95	289													
aag tca gag atg gag ttc att gaa agc aag cgc cct cgg cta gag ctg Lys Ser Glu Met Glu Phe Ile Glu Ser Lys Arg Pro Arg Leu Glu Leu 100 105 110	337													
ctg cct gac ccc ctg ctg cga ccg tca ccc ctg ctg gcc acg ggc cag Leu Pro Asp Pro Leu Leu Arg Pro Ser Pro Leu Leu Ala Thr Gly Gln 115 120 125	385													
cct gcg gga tct gaa gac ctc acc aag gac cgt agc ctg acg ggc aag Pro Ala Gly Ser Glu Asp Leu Thr Lys Asp Arg Ser Leu Thr Gly Lys 130 135 140	433													
ctg gaa ccg gtg tct ccc ccc agc ccc ccg cac act gac cct gag ctg Leu Glu Pro Val Ser Pro Pro Ser Pro Pro His Thr Asp Pro Glu Leu 145 150 155 160	481													
gag ctg gtg ccg cca cgg ctg tcc aag gag gag ctg atc cag aac atg Glu Leu Val Pro Pro Arg Leu Ser Lys Glu Glu Leu Ile Gln Asn Met 165 170 175	529													
gac cgc gtg gac cga gag atc acc atg gta gag cag cag atc tct aag Asp Arg Val Asp Arg Glu Ile Thr Met Val Glu Gln Gln Ile Ser Lys 180 185 190	577													
ctg aag aag aag cag caa cag ctg gag gag gat gcc aag ccg ccc Leu Lys Lys Lys Gln Gln Gln Leu Glu Glu Glu Ala Ala Lys Pro Pro 195 200 205	625													

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						tca Ser 215										673
						tac Tyr										721
						ggc Gly										769
tac Tyr	aac Asn	cag Gln	ccc Pro 260	tcc Ser	gac Asp	acc Thr	cgg Arg	cag Gln 265	tat Tyr	cat His	gag Glu	aac Asn	atc Ile 270	aaa Lys	ata Ile	817
aac Asn	cag Gln	gcg Ala 275	atg Met	cgg Arg	aag Lys	aag Lys	cta Leu 280	atc Ile	ttg Leu	tac Tyr	ttc Phe	aag Lys 285	agg Arg	agg Arg	aat Asn	865
cac His	gct Ala 290	cgg Arg	aaa Lys	caa Gln	tgg Trp	aag Lys 295	cag Gln	aag Lys	ttc Phe	tgc Cys	cag Gln 300	cgc Arg	tat Tyr	gac Asp	cag Gln	913
ctc Leu 305	atg Met	gag Glu	gcc Ala	ttg Leu	gaa Glu 310	aaa Lys	aag Lys	gtg Val	gag Glu	cgc Arg 315	atc Ile	gaa Glu	aac Asn	aac Asn	ccg Pro 320	961
						agc Ser										1009
ttc Phe	cct Pro	gag Glu	atc Ile 340	cgc Arg	aag Lys	cag Gln	cgc Arg	gag Glu 345	ctg Leu	cag Gln	gag Glu	cgc Arg	atg Met 350	cag Gln	agc Ser	1057
agg Arg	gtg Val	ggc Gly 355	cag Gln	cgg Arg	ggc Gly	agt Ser	ggg Gly 360	ctg Leu	tcc Ser	atg Met	tcg Ser	gcc Ala 365	gcc Ala	cgc Arg	agc Ser	1105
gag Glu	cac His 370	gag Glu	gtg Val	tca Ser	gag Glu	atc Ile 375	atc Ile	gat Asp	ggc Gly	ctc Leu	tca Ser 380	gag Glu	cag Gln	gag Glu	aac Asn	1153
ctg Leu 385	gag Glu	aag Lys	cag Gln	atg Met	cgc Arg 390	cag Gln	ctg Leu	gcc Ala	gtg Val	atc Ile 395	ccg Pro	ccc Pro	atg Met	ctg Leu	tac Tyr 400	1201
gac Asp	gct Ala	gac Asp	cag Gln	cag Gln 405	cgc Arg	atc Ile	aag Lys	ttc Phe	atc Ile 410	aac Asn	atg Met	aac Asn	gly aaa	ctt Leu 415	atg Met	1249
gcc Ala	gac Asp	ccc Pro	atg Met 420	aag Lys	gtg Val	tac Tyr	aaa Lys	gac Asp 425	cgc Arg	cag Gln	gtc Val	atg Met	aac Asn 430	atg Met	tgg Trp	1297
agt Ser	gag Glu	cag Gln 435	gag Glu	aag Lys	gag Glu	acc Thr	ttc Phe 440	cgg Arg	gag Glu	aag Lys	ttc Phe	atg Met 445	cag Gln	cat His	ccc Pro	1345

						gca Ala 455										1393
						tac Tyr										1441
						tat Tyr										1489
						cag Gln										1537
						gag Glu										1585
						aag Lys 535										1633
						gac Asp										1681
						aaa Lys										1729
aga Arg	cgc Arg	aaa Lys	ggc Gly 580	cgc Arg	atc Ile	acc Thr	cgc Arg	tca Ser 585	atg Met	gct Ala	aat Asn	gag Glu	gcc Ala 590	aac Asn	agc Ser	1777
						cag Gln										1825
ctg Leu	aat Asn 610	gag Glu	agt Ser	tct Ser	cgc Arg	tgg Trp 615	aca Thr	gaa Glu	gaa Glu	gaa Glu	atg Met 620	gaa Glu	aca Thr	gcc Ala	aag Lys	1873
aaa Lys 625	ggt Gly	ctc Leu	ctg Leu	gaa Glu	cac His 630	ggc Gly	cgc Arg	aac Asn	tgg Trp	tcg Ser 635	gcc Ala	atc Ile	gcc Ala	cgg Arg	atg Met 640	1921
gtg Val	ggc Gly	tcc Ser	aag Lys	act Thr 645	gtg Val	tcg Ser	cag Gln	tgt Cys	aag Lys 650	aac Asn	ttc Phe	tac Tyr	ttc Phe	aac Asn 655	tac Tyr	1969
aag Lys	aag Lys	agg Arg	cag Gln 660	aac Asn	ctc Leu	gat Asp	gag Glu	atc Ile 665	ttg Leu	cag Gln	cag Gln	cac His	aag Lys 670	ctg Leu	aag Lys	2017
atg Met	gag Glu	aag Lys 675	gag Glu	agg Arg	aac Asn	gcg Ala	cgg Arg 680	agg Arg	aag Lys	aag Lys	aag Lys	aaa Lys 685	gcg Ala	ccg Pro	gcg Ala	2065

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gcg Ala	gcc Ala 690	agc Ser	gag Glu	gag Glu	gct Ala	gca Ala 695	ttc Phe	ccg Pro	ccc Pro	gtg Val	gtg Val 700	gag Glu	gat Asp	gag Glu	gag Glu		2113
atg Met 705	gag Glu	gcg Ala	tcg Ser	ggc Gly	gtg Val 710	agc Ser	gga Gly	aat Asn	gag Glu	gag Glu 715	gag Glu	atg Met	gtg Val	gag Glu	gag Glu 720		2161
gct Ala	gaa Glu	gcc Ala	tta Leu	cat His 725	gcc Ala	tct Ser	Gly ggg	aat Asn	gag Glu 730	gtg Val	ccc Pro	aga Arg	G1A aaa	gaa Glu 735	tgc Cys	*	2209
agt Ser	ggc Gly	cca Pro	gcc Ala 740	act Thr	gtc Val	aac Asn	aac Asn	agc Ser 745	tca Ser	gac Asp	acc Thr	gag Glu	agc Ser 750	atc Ile	ccc Pro		2257
tct Ser	cct Pro	cac His 755	act Thr	gag Glu	gcc Ala	gcc Ala	aag Lys 760	gac Asp	aca Thr	ggg ggg	cag Gln	aat Asn 765	g1y 999	ccc Pro	aag Lys		2305
ccc Pro	cca Pro 770	gcc Ala	acc Thr	ctg Leu	ggc Gly	gcc Ala 775	gac Asp	gl <sup>à</sup> aaa	cca Pro	ccc Pro	cca Pro 780	ggc Gly	cca Pro	ccc Pro	acc Thr		2353
cca Pro 785	cca Pro	cgg Arg	agg Arg	aca Thr	tcc Ser 790	cgg Arg	gcc Ala	ccc Pro	att Ile	gag Glu 795	ccc Pro	acc Thr	ccg Pro	gcc Ala	tct Ser 800		2401
gaa Glu	gcc Ala	acc Thr	gga Gly	gcc Ala 805	cct Pro	acg Thr	ccc Pro	cca Pro	cca Pro 810	gca Ala	ccc Pro	cca Pro	tcg Ser	ccc Pro 815	tct Ser		2449
gca Ala	cct Pro	cct Pro	cct Pro 820	gtg Val	gtc Val	ccc Pro	aag Lys	gag Glu 825	gag Glu	aag Lys	gag Glu	gag Glu	gag Glu 830	acc Thr	gca Ala		2497
gca Ala	gcg Ala	ccc Pro 835	cca Pro	gtg Val	gag Glu	gag Glu	999 Gly 840	gag Glu	gag Glu	cag Gln	aag Lys	ccc Pro 845	ccc Pro	gcg Ala	gct Ala		2545
gag Glu	gag Glu 850	ctg Leu	gca Ala	gtg Val	gac Asp	aca Thr 855	Gly aaa	aag Lys	gcc Ala	gag Glu	gag Glu 860	ccc Pro	gtc Val	aag Lys	agc Ser		2593
gag Glu 865	tgc Cys	acg Thr	gag Glu	gaa Glu	gcc Ala 870	gag Glu	gag Glu	Gly 999	ccg Pro	gcc Ala 875	aag Lys	ggc	aag Lys	gac Asp	gcg Ala 880		2641
gag Glu	gcc Ala	gct Ala	gag Glu	gcc Ala 885	acg Thr	gcc Ala	gag Glu	gly aaa	gcg Ala 890	Leu	aag Lys	gca Ala	gag Glu	aag Lys 895	aag Lys		2689
gag Glu	ggc	Gly 999	agc Ser 900	ggc Gly	agg Arg	gcc Ala	acc Thr	act Thr 905	gcc Ala	aag Lys	agc Ser	tcg Ser	ggc Gly 910	Ala	ccc Pro		2737
cag Gln	gac Asp	agc Ser 915	Asp	tcc Ser	agt Ser	gct Ala	acc Thr 920	Cys	agt Ser	gca Ala	gac Asp	gag Glu 925	۷al	gat Asp	gag Glu		2785

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gcc Ala	gag Glu 930	ggc Gly	ggc Gly	gac Asp	aag Lys	aac Asn 935	cgg Arg	ctg Leu	ctg Leu	tcc Ser	cca Pro 940	agg Arg	ccc Pro	agc Ser	ctc Leu	2833
ctc Leu 945	acc Thr	ccg Pro	act Thr	ggc Gly	gac Asp 950	ccc Pro	cgg Arg	gcc Ala	aat Asn	gcc Ala 955	tca Ser	ccc Pro	cag Gln	aag Lys	cca Pro 960	2881
ctg Leu	gac Asp	ctg Leu	aag Lys	cag Gln 965	ctg Leu	aag Lys	cag Gln	cga Arg	gcg Ala 970	gct Ala	gcc Ala	atc Ile	ccc Pro	ccc Pro 975	atc Ile	2929
cag Gln	gtc Val	acc Thr	aaa Lys 980	gtc Val	cat His	gag Glu	ccc Pro	ccc Pro 985	cgg Arg	gag Glu	gac Asp	gca Ala	gct Ala 990	ccc Pro	acc Thr	2977
aag Lys	cca Pro	gct Ala 999	Pro	cca Pro	gcc Ala	cca Pro	ccg Pro 1000	Pro	ccg Pro	caa Gln	aac Asn	ctg Leu 1009	Gln	ccg Pro	gag Glu	3025
agc Ser	gac Asp 1010	Ala	cct Pro	cag Gln	cag Gln	cct Pro 1015	Gly	agc Ser	agc Ser	ccc Pro	cgg Arg 1020	Gly	aag Lys	agc Ser	agg Arg	3073
agc Ser 102	ccg Pro	gca Ala	ccc Pro	ccc Pro	gcc Ala 1030	Asp	aag Lys	gag Glu	gcc Ala	ttc Phe 1035	Ala	gcc Ala	gag Glu	gcc Ala	cag Gln 1040	3121
aag Lys	ctg Leu	cct Pro	gly ggg	gac Asp 104	Pro	cct Pro	tgc Cys	tgg Trp	act Thr 105	Ser	ggc Gly	ctg Leu	ccc Pro	ttc Phe 105	Pro	3169
gtg Val	ccc Pro	ccc Pro	cgt Arg 106	Glu	gtg Val	atc Ile	aag Lys	gcc Ala 106!	Ser	ccg Pro	cat His	gcc Ala	ccg Pro 107	Asp	ccc Pro	3217
tca Ser	gcc Ala	ttc Phe 107	Ser	tac Tyr	gct Ala	cca Pro	cct Pro 108	Gly	cac His	cca Pro	ctg Leu	ccc Pro 108	Leu	ggc Gly	ctc Leu	3265
cat His	gac Asp 109	Thr	gcc Ala	cgg Arg	ccc Pro	gtc Val 109	Leu	ccg Pro	cgc Arg	cca Pro	ccc Pro 110	Thr	atc Ile	tcc Ser	aac Asn	3313
ccg Pro 110	cct Pro 5	ccc Pro	ctc Leu	atc Ile	tcc Ser 111	Ser	gcc Ala	aag Lys	cac His	ccc Pro 111	Ser	gtc Val	ctc Leu	gag Glu	agg Arg 1120	3361
caa Gln	ata Ile	ggt Gly	gcc Ala	atc Ile 112	Ser	caa Gln	gga Gly	atg Met	tcg Ser 113	Val	cag Gln	ctc Leu	cac His	gtc Val 113	Pro	3409
tac Tyr	tca Ser	gag Glu	cat His 114	Ala	aag Lys	gcc Ala	ccg Pro	gtg Val 114	Gly	cct Pro	gtc Val	acc Thr	atg Met 115	Gly	ctg Leu	3457
ccc Pro	ctg Leu	ccc Pro 115	Met	gac Asp	ccc Pro	aaa Lys	aag Lys 116	Leu	gca Ala	ccc Pro	ttc Phe	agc Ser 116	Gly	gtg Val	aag Lys	3505

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		Gln					Gly				cca Pro 1180	Pro				3553
999 Gly 1185	Val	ccc Pro	aca Thr	gcc Ala	cag Gln 1190	Glu	gcg Ala	tcc Ser	gtg Val	ctg Leu 1195	aga Arg	Gly 999	aca Thr	gct Ala	ctg Leu 1200	3601
ggc Gly	tca Ser	gtt Val	ccg Pro	ggc Gly 1205	Gly	agc Ser	atc Ile	acc Thr	aaa Lys 1210	Gly	att Ile	ccc Pro	agc Ser	aca Thr 1215	Arg	3649
gtg Val	ccc Pro	tcg Ser	gac Asp 1220	Ser	gcc Ala	atc Ile	aca Thr	tac Tyr 1225	Arg	ggc Gly	tcc Ser	atc Ile	acc Thr 1230	His	ggc Gly	3697
acg Thr	cca Pro	gct Ala 1235	Asp	gtc Val	ctg Leu	tac Tyr	aag Lys 1240	Gly	acc Thr	atc Ile	acc Thr	agg Arg 1245	Ile	atc Ile	ggc Gly	3745
		Ser					Asp				gag Glu 1260	Asp				3793
aag Lys 1265	Gly	cac His	gtc Val	atc Ile	tac Tyr 1270	Glu	ggc Gly	aag Lys	aag Lys	ggc Gly 1275	cac His	gtc Val	ttg Leu	tcc Ser	tat Tyr 1280	3841
gag Glu	ggt Gly	ggc Gly	atg Met	tct Ser 1285	Val	acc Thr	cag Gln	tgc Cys	tcc Ser 1290	Lys	gag Glu	gac Asp	ggc Gly	aga Arg 129!	Ser	3889
agc Ser	tca Ser	gga Gly	ccc Pro 1300	Pro	cat His	gag Glu	acg Thr	gcc Ala 130	Ala	ccc Pro	aag Lys	cgc Arg	acc Thr 131	Tyr	gac Asp	3937
atg Met	atg Met	gag Glu 131!	Gly	cgc Arg	gtg Val	ggc Gly	aga Arg 132	Ala	atc Ile	tcc Ser	tca Ser	gcc Ala 132	Ser	atc Ile	gaa Glu	3985
ggt Gly	ctc Leu 133	Met	ggc Gly	cgt Arg	gcc Ala	atc Ile 133	Pro	ccg Pro	gag Glu	cga Arg	cac His 134	Ser	ccc Pro	cac His	cac His	4033
ctc Leu 134	Lys	gag Glu	cag Gln	cac His	cac His 135	Ile	cgc Arg	gly	tcc Ser	atc Ile 135	aca Thr 5	caa Gln	ggg Gly	atc Ile	cct Pro 1360	4081
cgg Arg	tcc Ser	tac Tyr	gtg Val	gag Glu 136	Ala	cag Gln	gag Glu	gac Asp	tac Tyr 137	Leu	cgt Arg	cgg Arg	gag Glu	gcc Ala 137	Lys	4129
ctc Leu	cta Leu	aag Lys	cgg Arg 138	Glu	ggc Gly	acg Thr	cct Pro	ccg Pro 138	Pro	cca Pro	ccg Pro	ccc Pro	tca Ser 139	Arg	gac Asp	4177
ctg Leu	acc Thr	gag Glu 139	Ala	tac Tyr	aag Lys	acg Thr	cag Gln 140	Ala	ctg Leu	ggc	ccc Pro	ctg Leu 140	Lys	ctg Leu	aag Lys	4225

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ccg Pro	gcc Ala 1410	His	gag Glu	ggc Gly	ctg Leu	gtg Val 1415	Ala	acg Thr	gtg Val	aag Lys	gag Glu 1420	Ala	ggc Gly	cgc Arg	tcc Ser	4273
atc Ile 1425	cat His	gag Glu	atc Ile	ccg Pro	cgc Arg 1430	Glu	gag Glu	ctg Leu	cgg Arg	cac His 1435	Thr	ccc Pro	gag Glu	ctg Leu	ccc Pro 1440	4321
ctg Leu	gcc Ala	ccg Pro	cgg Arg	ccg Pro 1445	Leu	aag Lys	gag Glu	ggc Gly	tcc Ser 1450	Ile	acg Thr	cag Gln	ggc Gly	acc Thr 1455	Pro	4369
ctc Leu	aag Lys	tac Tyr	gac Asp 1460	Thr	ggc Gly	gcg Ala	tcc Ser	acc Thr 1465	Thr	ggc Gly	tcc Ser	aaa Lys	aag Lys 1470	His	gac Asp	4417
gta Val	cgc Arg	tcc Ser 1475	Leu	atc Ile	ggc Gly	agc Ser	ccc Pro 1480	Gly	cgg Arg	acg Thr	ttc Phe	cca Pro 1485	Pro	gtg Val	cac His	4465
ccg Pro	ctg Leu 1490	Asp	gtg Val	atg Met	gcc Ala	gac Asp 1495	Ala	cgg Arg	gca Ala	ctg Leu	gaa Glu 1500	Arg	gcc Ala	tgc Cys	tac Tyr	4513
gag Glu 1509	gag Glu 5	agc Ser	ctg Leu	aag Lys	agc Ser 1510	Arg	cca Pro	ggg gly	acc Thr	gcc Ala 1515	Ser	agc Ser	tcg Ser	gly ggg	ggc Gly 1520	4561
tcc Ser	att Ile	gcg Ala	cgc Arg	ggc Gly 1525	Ala	ccg Pro	gtc Val	att Ile	gtg Val 1530	Pro	gag Glu	ctg Leu	ggt Gly	aag Lys 153!	Pro	4609
cgg Arg	cag Gln	agc Ser	ccc Pro 1540	Leu	acc Thr	tat Tyr	gag Glu	gac Asp 1545	His	gly ggg	gca Ala	ccc Pro	ttt Phe 155	Ala	ggc Gly	4657
cac His	ctc Leu	cca Pro 155!	Arg	ggt Gly	tcg Ser	ccc Pro	gtg Val 1560	Thr	atg Met	cgg Arg	gag Glu	ccc Pro 156	Thr	ccg Pro	cgc Arg	4705
ctg Leu	cag Gln 1570	Glu	ggc Gly	agc Ser	ctt Leu	tcg Ser 157	Ser	agc Ser	aag Lys	gca Ala	tcc Ser 1580	Gln	gac Asp	cga Arg	aag Lys	4753
ctg Leu 158	Thr	tcg Ser	acg Thr	cct Pro	cgt Arg 159	Glu	atc Ile	gcc Ala	aag Lys	tcc Ser 159	Pro	cac His	agc Ser	acc Thr	gtg Val 1600	4801
ccc Pro	gag Glu	cac His	cac His	cca Pro 160	His	ccc Pro	atc Ile	tcg Ser	ccc Pro 161	Tyr	gag Glu	cac His	ctg Leu	ctt Leu 161	Arg	4849
ggc Gly	gtg Val	agt Ser	ggc Gly 162	Val	gac Asp	ctg Leu	tat Tyr	cgc Arg 162	Ser	cac His	atc Ile	ccc Pro	ctg Leu 163	Ala	ttc Phe	4897
gac Asp	ccc Pro	acc Thr 163	Ser	ata Ile	ccc Pro	cgc Arg	ggc Gly 164	Ile	cct Pro	ctg Leu	gac Asp	gca Ala 164	Ala	gct Ala	gcc Ala	4945

tac tac ctg ccc cga cac ctg gcc ccc aac ccc acc tac ccg cac ctg Tyr Tyr Leu Pro Arg His Leu Ala Pro Asn Pro Thr Tyr Pro His Leu 1650 1655 1660	4993
tac cca ccc tac ctc atc cgc ggc tac ccc gac acg gcg gcg ctg gag Tyr Pro Pro Tyr Leu Ile Arg Gly Tyr Pro Asp Thr Ala Ala Leu Glu 1665 1670 1675 1680	5041
aac cgg cag acc atc atc aat gac tac atc acc tcg cag cag atg cac Asn Arg Gln Thr Ile Ile Asn Asp Tyr Ile Thr Ser Gln Gln Met His 1685 1690 1695	5089
cac aac acg gcc acc gcc atg gcc cag cga gct gat atg ctg agg ggc His Asn Thr Ala Thr Ala Met Ala Gln Arg Ala Asp Met Leu Arg Gly 1700 1705 1710	5137
ctc tcg ccc cgc gag tcc tcg ctg gca ctc aac tac gct gcg ggt ccc Leu Ser Pro Arg Glu Ser Ser Leu Ala Leu Asn Tyr Ala Ala Gly Pro 1715 1720 1725	5185
cga ggc atc atc gac ctg tcc caa gtg cca cac ctg cct gtg ctc gtg Arg Gly Ile Ile Asp Leu Ser Gln Val Pro His Leu Pro Val Leu Val 1730 1735 1740	5233
ccc ccg aca cca ggc acc cca gcc acc gcc atg gac cgc ctt gcc tacPro Pro Thr Pro Gly Thr Pro Ala Thr Ala Met Asp Arg Leu Ala Tyr174517501760	5281
ctc ccc acc gcg ccc cag ccc ttc agc agc cgc cac agc agc tcc cca Leu Pro Thr Ala Pro Gln Pro Phe Ser Ser Arg His Ser Ser Ser Pro 1765 1770 1775	5329
ctc tcc cca gga ggt cca aca cac ttg aca aaa cca acc acc acg tcc Leu Ser Pro Gly Gly Pro Thr His Leu Thr Lys Pro Thr Thr Ser 1780 1785 1790	5377
tcg tcc gag cgg gag cga gac cgg gat cga gag cgg gac cgg gat cgg Ser Ser Glu Arg Glu Arg Asp Arg Asp Arg Glu Arg Asp Arg Asp Arg 1795 1800 1805	5425
gag cgg gaa aag tcc atc ctc acg tcc acc acg acg gtg gag cac gca Glu Arg Glu Lys Ser Ile Leu Thr Ser Thr Thr Thr Val Glu His Ala 1810 1815 1820	5473
ccc atc tgg aga cct ggt aca gag cag agc agc ggc agc agc ggc agc Pro Ile Trp Arg Pro Gly Thr Glu Gln Ser Ser Gly Ser Ser Gly Ser 1825 1830 1835 1840	5521
agc ggc ggg ggt ggg ggc agc agc agc ccc gcc tcc cac tcc cat Ser Gly Gly Gly Gly Ser Ser Ser Arg Pro Ala Ser His Ser His 1845 1850 1855	5569
gcc cac cag cac tcg ccc atc tcc cct cgg acc cag gat gcc ctc cag Ala His Gln His Ser Pro Ile Ser Pro Arg Thr Gln Asp Ala Leu Gln 1860 1865 1870	5617
cag aga ccc agt gtg ctt cac aac aca ggc atg aag ggt atc atc acc Gln Arg Pro Ser Val Leu His Asn Thr Gly Met Lys Gly Ile Ile Thr 1875 1880 1885	5665

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		Glu					Thr				tcc Ser 1900	Thr				5713
tca Ser 1905	Pro	gtt Val	cgc Arg	cca Pro	gct Ala 1910	Ala	aca Thr	ttc Phe	cca Pro	cct Pro 1915	gcc Ala	acc Thr	cac His	tgc Cys	cca Pro 1920	5761
ctg Leu	ggc Gly	ggc Gly	acc Thr	ctc Leu 1925	Asp	ggg ggg	gtc Val	tac Tyr	cct Pro 1930	Thr	ctc Leu	atg Met	gag Glu	ccc Pro 1935	Val	5809
ttg Leu	ctg Leu	ccc Pro	aag Lys 1940	Glu	gcc Ala	ccc Pro	cgg Arg	gtc Val 1945	Ala	cgg Arg	cca Pro	gag Glu	cgg Arg 1950	Pro	cga Arg	5857
gca Ala	gac Asp	acc Thr 1955	Gly	cat His	gcc Ala	ttc Phe	ctc Leu 1960	Ala	aag Lys	ccc Pro	cca Pro	gcc Ala 1965	Arg	tcc Ser	gl <sup>à</sup> aaa	5905
ctg Leu	gag Glu 1970	Pro	gcc Ala	tcc Ser	tcc Ser	ccc Pro 1975	Ser	aag Lys	ggc Gly	tcg Ser	gag Glu 1980	Pro	cgg Arg	ccc Pro	cta Leu	5953
gtg Val 1985	Pro	cct Pro	gtc Val	tct Ser	ggc Gly 1990	His	gcc Ala	acc Thr	atc Ile	gcc Ala 1995	cgc Arg	acc Thr	cct Pro	gcg Ala	aag Lys 2000	6001
					His					Pro	ccg Pro				Ala	6049
tcg Ser	gcc Ala	tcg Ser	gac Asp 2020	Pro	cac His	cgg Arg	gaa Glu	aag Lys 2029	Thr	caa Gln	agt Ser	aaa Lys	ccc Pro 2030	Phe	tcc Ser	6097
atc Ile	cag Gln	gaa Glu 2035	Leu	gaa Glu	ctc Leu	cgt Arg	tct Ser 2040	Leu	ggt Gly	tac Tyr	cac His	ggc Gly 204!	Ser	agc Ser	tac Tyr	6145
agc Ser	ccc Pro 2050	Glu	gly aaa	gtg Val	gag Glu	ccc Pro 205	Val	agc Ser	cct Pro	gtg Val	agc Ser 2060	Ser	ccc Pro	agt Ser	ctg Leu	6193
acc Thr 206!	His	gac Asp	aag Lys	gly ggg	ctc Leu 2070	Pro	aag Lys	cac His	ctg Leu	gaa Glu 207!	gag Glu 5	ctc Leu	gac Asp	aag Lys	agc Ser 2080	6241
cac His	ctg Leu	gag Glu	Gly ggg	gag Glu 208	Leu	cgg Arg	ccc Pro	aag Lys	cag Gln 209	Pro	ggc Gly	ccc Pro	gtg Val	aag Lys 209	Leu	6289
gly ggc	gly aaa	gag Glu	gcc Ala 210	Ala	cac His	ctc Leu	cca Pro	cac His 210	Leu	cgg Arg	ccg Pro	ctg Leu	cct Pro 211	Glu	agc Ser	6337
cag Gln	ccc Pro	tcg Ser 211	Ser	agc Ser	ccg Pro	ctg Leu	ctc Leu 212	Gln	acc Thr	gcc Ala	cca Pro	999 Gly 212	Val	aaa Lys	ggt Gly	6385

cac cag cgg gtg gtc His Gln Arg Val Val 2130	acc ctg gcc ca Thr Leu Ala Gl 2135	g cac atc agt g n His Ile Ser G 2140	ag gtc atc aca lu Val Ile Thr	6433
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gcc ccc ctc tac tcc Ala Pro Leu Tyr Ser 216	Phe Pro Gly Al	c agc tgc ccc g a Ser Cys Pro V 2170	tc ctg gac ctc Val Leu Asp Leu 2175	6529
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1880

1875

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aca gtg gag cat of Thr Val Glu His i						Ser
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Glu Pro Glu Lys Pro Val Ser Pro Pro Pro Ile Glu Ser Lys His A 210 215 220										
Ser Leu Val Gln Ile Ile Tyr Asp Glu Asn Arg Lys Lys Ala Glu A										
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Tyr Asn Gln Pro Ser Asp Thr Arg Gln Tyr His Glu Asn Ile Lys I 260 265 270										
Asn Gln Ala Met Arg Lys Lys Leu Ile Leu Tyr Phe Lys Arg Arg A 275 280 285	sn.									
His Ala Arg Lys Gln Trp Glu Gln Arg Phe Cys Gln Arg Tyr Asp G 290 295 300	ln									
Leu Met Glu Ala Trp Glu Lys Lys Val Glu Arg Ile Glu Asn Asn F	ro,									

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36	
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Pro Gly Gly Gly Gly 2355			Ser Ser Arg
Lys Ala Lys Ser Pro	Ala Pro Gly Leu	Ala Ser Gly Asp	
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		_			ccc Pro 965	_			_	_	_					3287
					cac His											3335

acc atc agc agg atc gtc ggt gag gac agc cca agt cgc ctt gac cgg Thr Ile Ser Arg Ile Val Gly Glu Asp Ser Pro Ser Arg Leu Asp Arg 995 1000 1005	3383
gca cga gag gac acc ctg ccc aag ggc cat gtc atc tat gag ggc aag Ala Arg Glu Asp Thr Leu Pro Lys Gly His Val Ile Tyr Glu Gly Lys 1010 1015 1020	3431
aaa ggc cac gtc cta tcc tat gaa ggt ggt atg tcc gtg tca cag tgc Lys Gly His Val Leu Ser Tyr Glu Gly Gly Met Ser Val Ser Gln Cys 1025 1030 1035	3479
tct aag gag gat gga agg agc agc tcg ggc cca ccc cat gag act gcc Ser Lys Glu Asp Gly Arg Ser Ser Ser Gly Pro Pro His Glu Thr Ala 1040 1045 1050 1055	3527
gcc cct aaa cgc acc tat gac atg atg gag ggc cgt gta ggc agg act Ala Pro Lys Arg Thr Tyr Asp Met Met Glu Gly Arg Val Gly Arg Thr 1060 1065 1070	3575
gtc acc tca gcc agc ata gag gga ctc atg ggc cgc gcc atc cct gag Val Thr Ser Ala Ser Ile Glu Gly Leu Met Gly Arg Ala Ile Pro Glu 1075 1080 1085	3623
cag cac agc ccc cac ctc aag gag cag cat cac atc cga ggc tcc atc Gln His Ser Pro His Leu Lys Glu Gln His His Ile Arg Gly Ser Ile 1090 1095 1100	3671
acg caa ggc atc ccg agg tcc tat gtg gag gcg cag gag gac tac tta Thr Gln Gly Ile Pro Arg Ser Tyr Val Glu Ala Gln Glu Asp Tyr Leu 1105 1110 1115	3719
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ggt tcc atc acc cag ggc acc cca ctc aag tac gac tct ggg gca ccc Gly Ser Ile Thr Gln Gly Thr Pro Leu Lys Tyr Asp Ser Gly Ala Pro 1200 1205 1210 1215	4007
tcc act ggc acc aag aaa cac gac gtg cgc tcc atc atc ggc agc ccc Ser Thr Gly Thr Lys Lys His Asp Val Arg Ser Ile Ile Gly Ser Pro 1220 1225 1230	4055

								4	13						
ggc cgg Gly Arg			Pro					Leu					Asp		4103
cgg gca Arg Ala		Glu					Glu					Ser			4151
ggg acc Gly Thi 126	Ser					Gly					Gly				4199
gtc gtg Val Val 1280					Lys					Pro					4247
gac cac Asp His				Phe					Pro					Val	4295
acc acc			Pro	_				Gln					Leu		4343
agc aag Ser Lys		Ser					Leu					Arg			4391
gcc aag Ala Lys 134	s Ser	cca Pro	cac His	agc Ser	act Thr 1350	Val	ccc Pro	gag Glu	cac His	cac His 1355	Pro	cac His	ccc Pro	atc Ile	4439
tcc ccc Ser Pro 1360					Leu					Gly					4487
cgt ggt Arg Gly				Leu					Thr					Gly	4535
atc cct Ile Pro	ctg Leu	gaa Glu 139!	Ala	gca Ala	gcc Ala	gca Ala	gcc Ala 1400	Tyr	tac Tyr	ctg Leu	ccc Pro	cgg Arg 1409	His	ttg Leu	4583
gcc ccc Ala Pro		Pro					Leu					Leu			4631
ggc tac Gly Tyr 142	r Pro					Leu					Thr				4679
gac tac Asp Ty: 1440					Gln					Ala					4727
gcc caq Ala Gli				Met					Ser					Ser	4775

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caa gtg cca cac Gln Val Pro His 1490				o Gly Thr Pro	
gcc acc gcc atc Ala Thr Ala Ile 1505		Ala Tyr L			
ttc agc agc cgc Phe Ser Ser Arg 1520					
cac cta gct aaa His Leu Ala Lys		Thr Ser Se			
cgt gag cgg gaa Arg Glu Arg Glu 1559	Arg Asp Lys	tcc atc c Ser Ile L 1560	tc acg tct ac eu Thr Ser Th	c act aca gtg r Thr Thr Val 1565	5063
gag cat gca ccc Glu His Ala Pro 1570			hr Glu Gln Se		
ggg ggc agc agc Gly Gly Ser Ser 1585	cgc ccc gcc Arg Pro Ala 159	Ser His T	cc cac cag ca hr His Gln Hi 1595	c tcg ccc atc s Ser Pro Ile	5159
tcc ccc cgg acc Ser Pro Arg Thr 1600	cag gac gcc Gln Asp Ala 1605	ttg cag c Leu Gln G	ag agg ccc ag ln Arg Pro Se 1610	t gtg ctg cac r Val Leu His 161	
aac acg agc atg Asn Thr Ser Met	aag ggc gtg Lys Gly Val 1620	Val Thr S	cc gtg gaa co er Val Glu Pr .625	c ggc acg ccc o Gly Thr Pro 1630	5255
acg gtc ctg agg Thr Val Leu Arg 163	Trp Ala Arg	tcc acc t Ser Thr S 1640	cc acc tct to er Thr Ser Se	g cct gtc cgc r Pro Val Arg 1645	5303 T
cca gct gcc aca Pro Ala Ala Thr 1650			is Cys Pro Le		
ctt gaa ggg gtc Leu Glu Gly Val 1665		Leu Met G			
gag acc tct cgg Glu Thr Ser Arg 1680					7
cat gcc ttt ctt His Ala Phe Leu		Pro Gly A			

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aca gcc atc gcc Thr Ala Ile Ala 1730				His His Ala	5591
agt ccg gac ccg Ser Pro Asp Pro 1745		Thr Ser Al			5639
aag act caa agt Lys Thr Gln Ser 1760					5687
ctg ggt tac cac Leu Gly Tyr His		Gly Tyr Se			5735
atc agc ccg gtg Ile Ser Pro Val 179	Ser Ser Pro				5783
aaa cct ctg gaa Lys Pro Leu Glu 1810				y Glu Leu Arg	5831
cac aag cag cca His Lys Gln Pro 1825		Lys Leu Se			5879
cca cat ctg cgg Pro His Leu Arg 1840					5927
ctc cag act gcc Leu Gln Thr Ala		Lys Gly Hi			5975
gct cag cac atc Ala Gln His Ile 187	Ser Glu Val				6023
ccg cag cag ctc Pro Gln Gln Leu 1890				r Ser Phe Pro	6071
gga gcc agc tgc Gly Ala Ser Cys 1905		Asp Leu Ar			6119
tac ctc cca ccc Tyr Leu Pro Pro 1920					6167
agt gaa ggg ggc Ser Glu Gly Gly	aaa agg tcc Lys Arg Ser 1940	Pro Glu Pi	cc agc aaa aca ro Ser Lys Th: 945	a tcg gtc ctg r Ser Val Leu 1950	6215

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	gat gcc att gag Asp Ala Ile Gli 5			Gly Met
	cat gct cgg ago His Ala Arg Sen 197	r Thr Ala Tyr		
	ggc gag ccc agg Gly Glu Pro Arg 1990			
	cca acc ttc ttc Pro Thr Phe Phe 2005		ı Thr Glu Ser	
	tcg aag aag cag Ser Lys Lys Glr 2020			
	gag cca gaa tao Glu Pro Glu Tyi 5			Thr Glu
	ccc gcc atc act Pro Ala Ile Th 205	Gly Ala Gly		
	caa gaa cac gco Gln Glu His Ala 2070			
	gca ctc atg ggt Ala Leu Met Gly 2085		Gln Trp Glu	
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	tcg cca ggt gga Ser Pro Gly Gly 213	dly Gly Lys		
	cga aaa gcc aag Arg Lys Ala Lys 2150			<b>-</b>
	cct tct gtc tcc Pro Ser Val Ser 2165		Ser Glu Gly	
	cca ctc acc aac Pro Leu Thr Asn 2180			

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				Thr					Asn					agg Arg 5		6983
			Val					Pro					Ala	gca Ala		7031
		Pro					His					Glu		ccc Pro		7079
	Leu				cag Gln 2245	Tyr				_	Āsp	_		tga *		7124
gcag tccg aact acct gggg <210 <211 <212	gacaggtncgcaaggggggggggggggggggggggggggg	get get get get get tege tege tege tege	getga gtnea cecac cegar ancaa	actor actor ccco naago attor	cc ga at ct gg gg	agact cgtco ctggo gagto	tgagg catco cccto ggaca	g aag c aga g cga a gga	ggago agcto cagto aaggo	cccc ggca gacc ggag	tgag ttct ttac ggac	gtete geet cacte	gcc tgt cag	tgcgo ctaaa gggal	catgga cgtcca agcctt ttgttt angagg	7184 7244 7304 7364 7424 7465
		, D	, DCu	Lub												
<400 Met		Gly	Ser	Thr 5	Gln	Pro	Val	Ala	Gln 10	Thr	Trp	Arg	Ala	Ala 15	Glu	
	Arg	Tyr	Pro 20	-	His	Gly	Ile	Ser 25		Pro	Val	Gln	Ile 30	Ala	Arg	
Ser	His	Thr 35	Pro	Leu	Tyr	Asn	Gln 40		Ser	Asp	Thr	Arg 45	Gln	Tyr	His	
Glu	Asn 50	Ile	Lys	Ile	Asn	Gln 55	Ala	Met	Arg	Lys	Lys 60	Leu	Ile	Leu	Tyr	
Phe 65		Arg	Arg	Asn	His 70		Arg	Lys	Gln	Trp 75		Gln	Arg	Phe	Cys 80	
	Arg	Tyr	Asp	Gln 85		Met	Glu	Ala	Trp 90		Lys	Lys	Val	Glu 95		
Ile	Glu	Asn	Asn 100		Arg	Arg	Arg	Ala 105		Glu	Ser	Lys	Val 110	Arg	Glu	
Tyr	Tyr	Glu 115		Gln	Phe	Pro	Glu 120		Arg	Lys	Gln	Arg 125		Leu	Gln	
Glu	Arg 130		Gln	Ser	Arg	Val 135		Gln	Arg	Gly	Ser 140		Leu	Ser	Met	
Ser 145	Ala	Ala	Arg	Ser	Glu 150	His	Glu	Val	Ser	Glu 155	Ile	Ile	Asp	Gly	Leu 160	
	Glu	Gln	Glu			Glu	Lys	Gln			Gln	Leu	Ala	Val		
Pro	Pro	Met	Leu 180	165 Tyr	Asp	Ala	Asp	Gln 185	170 Gln	Arg	Ile	Lys	Phe	175 Ile	Asn	
Met	Asn	Gly 195		Met	Asp	Asp	Pro 200		Lys	Val	Tyr	-		Arg	Gln	
Val	Thr 210		Met	Trp	Ser			Glu	Arg	Asp		205 Phe	Arg	Glu	Lys	
Phe		Gln	His	Pro	Lys	215 Asn	Phe	Gly	Leu	Ile	220 Ala	Ser	Phe	Leu	Glu	
225	T	mh	₹7~ <sup>7</sup>	7A 7 -	230	C	77-7	T 0	(T)	235	П	т о	mp	T	240	
Ara	LVS	ınr	val	AIA	(+ 111	UVS	val	uell	IVY	TVY	IVY	ueu	nnr	Lvs	LVS	

Arg Lys Thr Val Ala Glu Cys Val Leu Tyr Tyr Tyr Leu Thr Lys Lys

245 250 Asn Glu Asn Tyr Lys Ser Leu Val Arg Arg Ser Tyr Arg Arg Arg Gly 260 265 Met Ala Arg Ser Ser Gln Glu Glu Lys Glu Lys Glu Lys Glu Lys 300 Glu Ala Asp Lys Glu Glu Glu Lys Gln Asp Ala Glu Asn Glu Lys Glu 310 315 Glu Leu Ser Lys Glu Lys Thr Asp Asp Thr Ser Gly Glu Asp Asn Asp 330 Glu Lys Glu Ala Val Ala Ser Lys Gly Arg Lys Thr Ala Asn Ser Gln 345 Gly Arg Arg Lys Gly Arg Ile Thr Arg Ser Met Ala Asn Glu Ala Asn 360 His Glu Glu Thr Ala Thr Pro Gln Gln Ser Ser Glu Leu Ala Ser Met 375 380 Glu Met Asn Glu Ser Ser Arg Trp Thr Glu Glu Glu Met Glu Thr Ala 390 395 Lys Lys Gly Leu Leu Glu His Gly Arg Asn Trp Ser Ala Ile Ala Arg 405 410 Met Val Gly Ser Lys Thr Val Ser Gln Cys Lys Asn Phe Tyr Phe Asn 420 425 Tyr Lys Lys Arg Gln Asn Leu Asp Glu Ile Leu Gln Gln His Lys Leu 440 Lys Met Glu Lys Glu Arg Asn Ala Arg Arg Lys Lys Lys Thr Pro 455 Ala Ala Ser Glu Glu Thr Ala Phe Pro Pro Ala Ala Glu Asp Glu 470 475 Glu Met Glu Ala Ser Gly Ala Ser Ala Asn Glu Glu Glu Leu Ala Glu 490 Glu Ala Glu Ala Ser Gln Ala Ser Gly Asn Glu Val Pro Arg Val Gly 505 Glu Cys Ser Gly Pro Ala Ala Val Asn Asn Ser Ser Asp Thr Glu Ser 520 Val Pro Ser Pro Arg Ser Glu Ala Thr Lys Asp Thr Gly Pro Lys Pro 535 Thr Gly Thr Glu Ala Leu Pro Ala Ala Thr Gln Pro Pro Val Pro Pro 550 555 Pro Glu Glu Pro Ala Val Ala Pro Ala Glu Pro Ser Pro Val Pro Asp 570 Ala Ser Gly Pro Pro Ser Pro Glu Pro Ser His His Leu Pro His Pro 585 Arg Leu Leu Trp Thr Arg Met Asn Lys Lys Pro Arg Leu Leu Gln Leu 600 Pro Arg Gln Arg Met Pro Arg Ser Arg Ser Leu Arg Pro Arg Arg Ser 615 620 Met Trp Glu Lys Pro Glu Glu Pro Glu Ala Ser Glu Lys Pro Pro Lys 630 635 Ser Val Lys Ser Asp His Lys Lys Glu Thr Glu Glu Glu Pro Glu Asp 645 650 Lys Ala Lys Gly Thr Glu Ala Ile Glu Thr Val Ser Glu Ala Pro Leu 665 Lys Val Glu Lys Ala Gly Ser Lys Ala Ala Val Thr Lys Gly Ser Ser 680 685 Ser Gly Ala Thr Gln Asp Ser Asp Ser Ser Ala Thr Cys Ser Ala Asp 695 700 Glu Val Asp Glu Pro Glu Gly Gly Asp Lys Gly Arg Leu Leu Ser Pro 710 715 Arg Pro Ser Leu Leu Thr Pro Ala Gly Asp Pro Arg Ala Ser Thr Ser

725 730 Pro Gln Lys Pro Leu Asp Leu Lys Gln Leu Lys Gln Arg Ala Ala 740 745 Ile Pro Pro Ile Val Thr Lys Val His Glu Pro Pro Arg Glu Asp Thr 760 Val Pro Pro Lys Pro Val Pro Pro Val Pro Pro Pro Thr Gln His Leu 775 Gln Pro Glu Gly Asp Val Ser Gln Gln Ser Gly Gly Ser Pro Arg Gly 795 790 Lys Ser Arg Ser Pro Val Pro Pro Ala Glu Lys Glu Ala Glu Lys Pro 810 Ala Phe Pro Ala Phe Pro Thr Glu Gly Pro Lys Leu Pro Thr Glu 825 820 Pro Pro Arg Trp Ser Ser Gly Leu Pro Phe Pro Ile Pro Pro Arg Glu 840 Val Ile Lys Thr Ser Pro His Ala Ala Asp Pro Ser Ala Phe Ser Tyr 855 860 Thr Pro Pro Gly His Pro Leu Pro Leu Gly Leu His Asp Ser Ala Arg 870 875 Pro Val Leu Pro Arg Pro Pro Ile Ser Asn Pro Pro Pro Leu Ile Ser 885 890 Ser Ala Lys His Pro Gly Val Leu Glu Arg Gln Leu Gly Ala Ile Ser 900 905 Gln Gln Gly Met Ser Val Gln Leu Arg Val Pro His Ser Glu His Ala 920 Lys Ala Pro Met Gly Pro Leu Thr Met Gly Leu Pro Leu Ala Val Asp 935 Pro Lys Lys Leu Gly Thr Ala Leu Gly Ser Ala Thr Ser Gly Ser Ile 950 955 Thr Lys Gly Leu Pro Ser Thr Arg Ala Ala Asp Gly Pro Ser Tyr Arg 965 970 Gly Ser Ile Thr His Gly Thr Pro Ala Asp Val Leu Tyr Lys Gly Thr 985 Ile Ser Arg Ile Val Gly Glu Asp Ser Pro Ser Arg Leu Asp Arg Ala 1000 1005 Arg Glu Asp Thr Leu Pro Lys Gly His Val Ile Tyr Glu Gly Lys Lys 1010 1015 1020 Gly His Val Leu Ser Tyr Glu Gly Gly Met Ser Val Ser Gln Cys Ser 1030 1035 Lys Glu Asp Gly Arg Ser Ser Ser Gly Pro Pro His Glu Thr Ala Ala 1045 1050 1055 Pro Lys Arg Thr Tyr Asp Met Met Glu Gly Arg Val Gly Arg Thr Val 1060 1065 Thr Ser Ala Ser Ile Glu Gly Leu Met Gly Arg Ala Ile Pro Glu Gln 1080 1085 His Ser Pro His Leu Lys Glu Gln His His Ile Arg Gly Ser Ile Thr 1090 1095 1100 Gln Gly Ile Pro Arg Ser Tyr Val Glu Ala Gln Glu Asp Tyr Leu Arg 1110 1115 1120 Arg Glu Ala Lys Leu Leu Lys Arg Glu Gly Thr Pro Pro Pro Pro 1130 1135 1125 Pro Pro Arg Asp Leu Thr Glu Thr Tyr Lys Pro Arg Pro Leu Asp Pro 1140 1145 1150 Leu Gly Pro Leu Lys Leu Lys Pro Thr His Glu Gly Val Val Ala Thr 1155 1160 1165 Val Lys Glu Ala Gly Arg Ser Ile His Glu Ile Pro Arg Glu Glu Leu 1170 1175 1180 Arg Arg Thr Pro Glu Leu Pro Leu Ala Pro Arg Pro Leu Lys Glu Gly 1190 1195 Ser Ile Thr Gln Gly Thr Pro Leu Lys Tyr Asp Ser Gly Ala Pro Ser

1205 1210 Thr Gly Thr Lys Lys His Asp Val Arg Ser Ile Ile Gly Ser Pro Gly 1220 1225 Arg Pro Phe Pro Ala Leu His Pro Leu Asp Ile Met Ala Asp Ala Arg 1235 1240 1245 Ala Leu Glu Arg Ala Cys Tyr Glu Glu Ser Leu Lys Ser Arg Ser Gly 1250 1255 1260 Thr Ser Ser Gly Ala Gly Gly Ser Ile Thr Arg Gly Ala Pro Val Val 1270 1275 Val Pro Glu Leu Gly Lys Pro Arg Gln Ser Pro Leu Thr Tyr Glu Asp 1285 1290 1295 His Gly Ala Pro Phe Thr Ser His Leu Pro Arg Gly Ser Pro Val Thr 1305 1310 1300 Thr Arg Glu Pro Thr Pro Arg Leu Gln Glu Gly Ser Leu Leu Ser Ser 1315 1320 1325 Lys Ala Ser Gln Asp Arg Lys Leu Thr Ser Thr Pro Arg Glu Ile Ala 1335 1340 Lys Ser Pro His Ser Thr Val Pro Glu His His Pro His Pro Ile Ser 1350 1355 1360 Pro Tyr Glu His Leu Leu Arg Gly Val Thr Gly Val Asp Leu Tyr Arg 1365 1370 1375 Gly His Ile Pro Leu Ala Phe Asp Pro Thr Ser Ile Pro Arg Gly Ile 1380 1385 1390 Pro Leu Glu Ala Ala Ala Ala Tyr Tyr Leu Pro Arg His Leu Ala 1395 1400 1405 Pro Ser Pro Thr Tyr Pro His Leu Tyr Pro Pro Tyr Leu Ile Arg Gly 1410 1415 1420 Tyr Pro Asp Thr Ala Ala Leu Glu Asn Arg Gln Thr Ile Ile Asn Asp 1430 1435 Tyr Ile Thr Ser Gln Gln Met His His Asn Ala Ala Ser Ala Met Ala 1450 Gln Arg Ala Asp Met Leu Arg Gly Leu Ser Pro Arg Glu Ser Ser Leu 1460 1465 Ala Leu Asn Tyr Ala Ala Gly Pro Arg Gly Ile Ile Asp Leu Ser Gln 1475 1480 Val Pro His Leu Pro Val Leu Val Pro Pro Thr Pro Gly Thr Pro Ala 1495 1500 Thr Ala Ile Asp Arg Leu Ala Tyr Leu Pro Thr Ala Pro Pro Pro Phe 1510 1515 Ser Ser Arg His Ser Ser Ser Pro Leu Ser Pro Gly Gly Pro Thr His 1525 1530 Leu Ala Lys Pro Thr Ala Thr Ser Ser Ser Glu Arg Glu Arg 1540 1545 Glu Arg Glu Arg Asp Lys Ser Ile Leu Thr Ser Thr Thr Thr Val Glu 1555 1560 1565 His Ala Pro Ile Trp Arg Pro Gly Thr Glu Gln Ser Ser Gly Ala Gly 1575 1580 Gly Ser Ser Arg Pro Ala Ser His Thr His Gln His Ser Pro Ile Ser 1590 1595 1600 Pro Arg Thr Gln Asp Ala Leu Gln Gln Arg Pro Ser Val Leu His Asn 1605 1610 1615 Thr Ser Met Lys Gly Val Val Thr Ser Val Glu Pro Gly Thr Pro Thr 1620 1625 1630 Val Leu Arg Trp Ala Arg Ser Thr Ser Thr Ser Ser Pro Val Arg Pro 1640 1645 1635 Ala Ala Thr Phe Pro Pro Ala Thr His Cys Pro Leu Gly Gly Thr Leu 1650 1655 1660 Glu Gly Val Tyr Pro Thr Leu Met Glu Pro Val Leu Leu Pro Lys Glu 1670 1675 Thr Ser Arg Val Ala Arg Pro Glu Arg Ala Arg Val Asp Ala Gly His

1685 1690 Ala Phe Leu Thr Lys Pro Pro Gly Arg Glu Pro Ala Ser Ser Pro Ser 1700 1705 1710 Lys Ser Ser Glu Pro Arg Ser Leu Ala Pro Pro Ser Ser Ser His Thr 1715 1720 1725 Ala Ile Ala Arg Thr Pro Ala Lys Asn Leu Ala Pro His His Ala Ser 1735 1740 Pro Asp Pro Pro Ala Pro Thr Ser Ala Ser Asp Leu His Arg Glu Lys 1750 1755 Thr Gln Ser Lys Pro Phe Ser Ile Gln Glu Leu Glu Leu Arg Ser Leu 1765 1770 1775 Gly Tyr His Ser Gly Ala Gly Tyr Ser Pro Asp Gly Val Glu Pro Ile 1780 1785 1790 Ser Pro Val Ser Ser Pro Ser Leu Thr His Asp Lys Gly Leu Ser Lys 1795 1800 1805 Pro Leu Glu Glu Leu Glu Lys Ser His Leu Glu Gly Glu Leu Arg His 1815 1820 Lys Gln Pro Gly Pro Met Lys Leu Ser Ala Glu Ala Ala His Leu Pro 1830 1835 1840 His Leu Arg Pro Leu Pro Glu Ser Gln Pro Ser Ser Pro Leu Leu 1845 1850 1855 Gln Thr Ala Pro Gly Ile Lys Gly His Gln Arg Val Val Thr Leu Ala 1865 1870 Gln His Ile Ser Glu Val Ile Thr Gln Asp Tyr Thr Arg His His Pro 1875 1880 1885 Gln Gln Leu Ser Gly Pro Leu Pro Ala Pro Leu Tyr Ser Phe Pro Gly 1890 1895 1900 Ala Ser Cys Pro Val Leu Asp Leu Arg Arg Pro Pro Ser Asp Leu Tyr 1910 1915 1920 Leu Pro Pro Pro Asp His Gly Thr Pro Ala Arg Gly Ser Pro His Ser 1925 1930 1935 Glu Gly Gly Lys Arg Ser Pro Glu Pro Ser Lys Thr Ser Val Leu Gly 1940 1945 Ser Ser Glu Asp Ala Ile Glu Pro Val Ser Pro Pro Glu Gly Met Thr 1955 1960 1965 Glu Pro Gly His Ala Arg Ser Thr Ala Tyr Pro Leu Leu Tyr Arg Asp 1970 1975 1980 Gly Glu Gln Gly Glu Pro Arg Met Gly Leu Glu Ser Pro Gly Asn Thr 1990 1995 Ser Gln Pro Pro Thr Phe Phe Ser Lys Leu Thr Glu Ser Asn Ser Ala 2005 2010 Met Val Lys Ser Lys Lys Gln Glu Ile Asn Lys Lys Leu Asn Thr His 2025 Asn Arg Asn Glu Pro Glu Tyr Asn Ile Gly Gln Pro Gly Thr Glu Ile 2035 2040 2045 Phe Asn Met Pro Ala Ile Thr Gly Ala Gly Leu Met Thr Cys Arg Ser 2055 2060 Gln Ala Val Gln Glu His Ala Ser Thr Asn Met Gly Leu Glu Ala Ile 2070 2075 Ile Arg Lys Ala Leu Met Gly Lys Tyr Asp Gln Trp Glu Glu Pro Pro 2085 2090 Pro Leu Gly Ala Asn Ala Phe Asn Pro Leu Asn Ala Ser Ala Ser Leu 2100 2105 Pro Ala Ala Ala Met Pro Ile Thr Thr Ala Asp Gly Arg Ser Asp His 2115 2120 2125 Ala Leu Thr Ser Pro Gly Gly Gly Lys Ala Lys Val Ser Gly Arg 2135 2140 Pro Ser Ser Arg Lys Ala Lys Ser Pro Ala Pro Gly Leu Ala Ser Gly 2150 2155 Asp Arg Pro Pro Ser Val Ser Ser Val His Ser Glu Gly Asp Cys Asn

			52		
	2165		2170		2175
Arg Arg Thr Pr 21	o Leu Thr A: 80	n Arg Val 218!		Asp Arg Pro 219	
Ala Gly Ser Th	r Pro Phe P	o Tyr Asn 2200	Pro Leu	Ile Met Arg 2205	Leu Gln
Ala Gly Val Me 2210		o Pro Pro	Pro Gly	Leu Ala Ala 2220	Gly Ser
Gly Pro Leu Al 2225	a Gly Pro H: 2230	s His Ala	Trp Asp	Glu Glu Pro	Lys Pro 2240
Leu Leu Cys Se		lu Thr Leu		_	2210
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agaaacatga ttg					
atg tca agt tc Met Ser Ser Se	r Gly Tyr Pı		Gln Gly		Thr Glu
1	5		10		15
caa agt cgt ta Gln Ser Arg Ty 2	r Pro Pro Hi				
cgc cac cag ca Arg His Gln Gl 35		_	_	_	
gaa gtg agt ca Glu Val Ser Gl 50	n Ala Ser Gl				
ctt cga agg cg		•	_		_
Leu Arg Arg Ar 65	g Pro Ser Le 70	u Leu Ser	Glu Phe 75	His Pro Gly	Ser Asp 80
agg cct caa ga Arg Pro Gln Gl					
tcc cca gtg ga Ser Pro Val As	_				<b>J J</b>
10		105		110	
cag gtt tct ga Gln Val Ser As 115					
tta gtg cac cc Leu Val His Pr					

	gat Asp		-						_	_						720
	ggg Gly															768
	aag Lys															816
_	aaa Lys	_	_	_	_						_				_	864
	gaa Glu 210	_		_	_							-				912
	cct Pro				-			_	_		_					960
_	gag Glu					_	_	_	-					_		1008
	ggc Gly			_	_	_		_			_			_		1056
	gtg Val															1104
	att Ile 290															1152
caa Gln 305	aaa Lys	atc Ile	tgc Cys	cag Gln	cgt Arg 310	tat Tyr	gat Asp	cag Gln	ctc Leu	atg Met 315	gag Glu	gca Ala	tgg Trp	gag Glu	aaa Lys 320	1200
	gtg Val															1248
	aca Thr		_				_	_			_		_			1296
_	gaa Glu	_		_	_		_	_	_		-			_		1344
	tca Ser	_			_		_						_			1392

gat ggg ctc tct gag cag gag aat aat gag aaa caa atg cgg cag ctc

Asp Gly Leu Ser Glu Gln Glu Asn Asn Glu Lys Gln Met Arg Gln Leu

tct gtg att cca cct atg atg ttt gat gca gaa caa aga cga gtc aag

Ser Val Ile Pro Pro Met Met Phe Asp Ala Glu Gln Arg Arg Val Lys

				_	aat Asn			_		_		_					1536
	_		_		atg Met		_			_		_	_	_			1584
	_	_	_		atc Ile	-									_		1632
2 2300 mg. 2 2000 mg. 2 2000 mg.		_			aag Lys	_	_		_	_	_	_					1680
The day was the fact that the	_				gag Glu 485				_		_	_					1728
the the the training of training of the training of the training of the training of tr		_	_		aga Arg		_			_	_				-	_	1776
		_	_	_	aaa Lys	_		_		_	_			_			1824
	_	_	_	_	aaa Lys	_	_		_		_	_			_		1872
		_			aag Lys	_	_	_	_		_			_	_	_	1920
				_	gag Glu 565		_					_	_		_		1968
	_	_		_	cgt Arg	_							_			_	2016
	_	_	_	_	agt Ser	_	_	_	_		_		_				2064

cca cct ctg cca ccg cca cca gaa ccc att tct aca gag cct gtg gag

Pro Pro Leu Pro Pro Pro Pro Glu Pro Ile Ser Thr Glu Pro Val Glu

		_	tgg Trp		_	_										2160
			ggt Gly													2208
	_	_	gct Ala 660		_										_	2256
			gac Asp													2304
aaa Lys	cct Pro 690	cgt Arg	gaa Glu	gag Glu	cga Arg	gat Asp 695	gtg Val	tct Ser	caa Gln	tgt Cys	gaa Glu 700	agt Ser	gtc Val	gct Ala	tcc Ser	2352
			gct Ala													2400
			gaa Glu													2448
			aat Asn 740													2496
ctt Leu	gag Glu	ccc Pro 755	acc Thr	acg Thr	gaa Glu	act Thr	gca Ala 760	ccc Pro	agt Ser	aca Thr	tct Ser	ccc Pro 765	tcc Ser	tta Leu	gca Ala	2544
			aca Thr													2592
			atc Ile												cag Gln 800	2640
			agt Ser													2688
			gac Asp 820													2736
			gtt Val													2784
gcc Ala	agt Ser	gag Glu	aag Lys	gtg Val	gaa Glu	cct Pro	aga Arg	gat Asp	gaa Glu	gat Asp	ttg Leu	gtg Val	gta Val	gct Ala	cag Gln	2832

	ata Ile		_								_		_		_	2880
	acg Thr															2928
	atg Met			_	_		-			_						2976
	ata Ile		_			_						_	_	~		3024
	ctt Leu 930										_	_		_		3072
	tgt Cys															3120
	cga Arg															3168
	cag Gln															3216
	tgt Cys		Thr		_	_		Asn	_			_	Val		_	3264
	gct Ala 1010	Pro			_		Thr				_	Gly	_			3312
_	aca Thr		_			Arg		_			Leu		_			3360
	acc Thr				Ser					Phe					Ser	3408
	tca Ser	_		Thr					Leu					Gln	_	3456
	tac Tyr		Gln	_			_	Pro		_			Ile			3504
	ctg Leu				_	_		_			_		_			3552

	Lys					Ser					aac Asn					3600
					Ala					Val	gtc Val				Ala	3648
				Glu					Arg		act Thr			Ser		3696
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		Āla					Gly				gag Glu 1180	Ala				3792
	Ser					Pro					agt Ser					3840
					Ser					Ile	tat Tyr				Ser	3888
				Ser					Lys		gcc Ala			Gly		3936
			Arg					Ile			aag Lys		Ser			3984
tca Ser	gtg Val 1250	Glu	gga Gly	aat Asn	ata Ile	aag Lys 125!	Gln	Gly ggg	atg Met	tca Ser	atg Met 1260	Arg	gag Glu	tct Ser	cct Pro	4032
	Ser					Gly					gca Ala 5					4080
agt Ser	cct Pro	cat His	tct Ser	gac Asp 128	Leu	aaa Lys	gaa Glu	agg Arg	act Thr 129	Val	ttg Leu	tct Ser	ggc Gly	tcc Ser 129	Ile	4128
atg Met	cag Gln	gly 999	aca Thr 130	Pro	aga Arg	gca Ala	aca Thr	act Thr 130	Glu	agc Ser	ttt Phe	gaa Glu	gat Asp 131	Gly	ctt Leu	4176
aaa Lys	tat Tyr	ccc Pro 131	Lys	caa Gln	att Ile	aaa Lys	agg Arg 132	Glu	agt Ser	cct Pro	ccc Pro	ata Ile 132	Arg	gca Ala	ttt Phe	4224
gaa Glu	ggt Gly	gcc Ala	att Ile	acc Thr	aaa Lys	gga Gly	aaa Lys	cca Pro	tat Tyr	gat Asp	ggc Gly	atc Ile	acc Thr	acc Thr	atc Ile	4272

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act cag gaa agt cgg aaa act cca gaa gtg gtc cag agc aca cgg ccg Thr Gln Glu Ser Arg Lys Thr Pro Glu Val Val Gln Ser Thr Arg Pro 1365 1370 1375	4368
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ggg cct agc aaa cta tcc cgt gga atg cct ccg ctg gaa att gtg cca Gly Pro Ser Lys Leu Ser Arg Gly Met Pro Pro Leu Glu Ile Val Pro 1410 1415 1420	4512
gag aac ata aaa gtg gta gaa cgg gga aaa tat gag gat gtg aaa gca Glu Asn Ile Lys Val Val Glu Arg Gly Lys Tyr Glu Asp Val Lys Ala 1425 1430 1435 1440	4560
ggc gag acc gtg cgt tcc cgg cac acg tca gtg gta agc tct ggc ccc Gly Glu Thr Val Arg Ser Arg His Thr Ser Val Val Ser Ser Gly Pro 1445 1450 1455	4608
tcc gtt ctt agg tcc aca ctg cat gaa gct ccc aaa gca caa ctg agc Ser Val Leu Arg Ser Thr Leu His Glu Ala Pro Lys Ala Gln Leu Ser 1460 1465 1470	4656
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aga gga ctc tcc cca aga gag cag cca ctg ggt ctc cca tac cca gca Arg Gly Leu Ser Pro Arg Glu Gln Pro Leu Gly Leu Pro Tyr Pro Ala 1635 1640 1645	5184
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cct cat cca ggg gga aca agc act cct ccc atg gac aga atc act tat Pro His Pro Gly Gly Thr Ser Thr Pro Pro Met Asp Arg Ile Thr Tyr 1665 1670 1675 1680	5280
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gtg gag aag aga tot gtt cag tgt tta tac act tot toa gcc Val Glu Lys Arg Ser Val Gln Cys Leu Tyr Thr Ser Ser Ala 1875 1880 1885	ttt cca 5904 Phe Pro
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tct tca cag gca gag gga atg ggg caa gtg ccc agg acc cat Ser Ser Gln Ala Glu Gly Met Gly Gln Val Pro Arg Thr His 2035 2040 2045	cgg ctg 6384 Arg Leu
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Arg His Gln Glu Phe Ala Val Pro Asp Tyr Arg Ser Ser His Leu 35 40 45	
Glu Val Ser Gln Ala Ser Gln Leu Leu Gln Gln Gln Gln Gln Gln	
50 55 60 Leu Arg Arg Pro Ser Leu Leu Ser Glu Phe His Pro Gly Ser Asp	

75 70 65 Arg Pro Gln Glu Arg Arg Thr Ser Tyr Glu Pro Phe His Pro Gly Pro 90 85 Ser Pro Val Asp His Asp Ser Leu Glu Ser Lys Arg Pro Arg Leu Glu 105 100 Gln Val Ser Asp Ser His Phe Gln Arg Val Ser Ala Ala Val Leu Pro 120 Leu Val His Pro Leu Pro Glu Gly Leu Arg Ala Ser Ala Asp Ala Lys 140 135 Lys Asp Pro Ala Phe Gly Gly Lys His Glu Ala Pro Ser Ser Pro Ile 155 150 Ser Gly Gln Pro Cys Gly Asp Asp Gln Asn Ala Ser Pro Ser Lys Leu 170 Ser Lys Glu Glu Leu Ile Gln Ser Met Asp Arg Val Asp Arg Glu Ile 185 Ala Lys Val Glu Gln Gln Ile Leu Lys Leu Lys Lys Lys Gln Gln Gln 200 205 Leu Glu Glu Glu Ala Ala Lys Pro Pro Glu Pro Glu Lys Pro Val Ser 215 Pro Pro Pro Val Glu Gln Lys His Arg Ser Ile Val Gln Ile Ile Tyr 230 235 Asp Glu Asn Arg Lys Lys Ala Glu Glu Ala His Lys Ile Phe Glu Gly 245 Leu Gly Pro Lys Val Glu Leu Pro Leu Tyr Asn Gln Pro Ser Asp Thr 265 260 Lys Val Tyr His Glu Asn Ile Lys Thr Asn Gln Val Met Arg Lys Lys 280 275 Leu Ile Leu Phe Phe Lys Arg Arg Asn His Ala Arg Lys Gln Arg Glu 295 Gln Lys Ile Cys Gln Arg Tyr Asp Gln Leu Met Glu Ala Trp Glu Lys 315 310 Lys Val Asp Arg Ile Glu Asn Asn Pro Arg Arg Lys Ala Lys Glu Ser 330 325 Lys Thr Arg Glu Tyr Tyr Glu Lys Gln Phe Pro Glu Ile Arg Lys Gln 345 340 Arg Glu Gln Gln Glu Arg Phe Gln Arg Val Gly Gln Arg Gly Ala Gly 360 Leu Ser Ala Thr Ile Ala Arg Ser Glu His Glu Ile Ser Glu Ile Ile 380 375 Asp Gly Leu Ser Glu Gln Glu Asn Asn Glu Lys Gln Met Arg Gln Leu 395 390 Ser Val Ile Pro Pro Met Met Phe Asp Ala Glu Gln Arg Arg Val Lys 410 405 Phe Ile Asn Met Asn Gly Leu Met Glu Asp Pro Met Lys Val Tyr Lys 425 420 Asp Arg Gln Phe Met Asn Val Trp Thr Asp His Glu Lys Glu Ile Phe 440 435 Lys Asp Lys Phe Ile Gln His Pro Lys Asn Phe Gly Leu Ile Ala Ser 455 460 Tyr Leu Glu Arg Lys Ser Val Pro Asp Cys Val Leu Tyr Tyr Tyr Leu 475 470 Thr Lys Lys Asn Glu Asn Tyr Lys Ala Leu Val Arg Arg Asn Tyr Gly 490 485 Lys Arg Arg Gly Arg Asn Gln Gln Ile Ala Arg Pro Ser Gln Glu Glu 505 500 Lys Val Glu Glu Lys Glu Glu Asp Lys Ala Glu Lys Thr Glu Lys Lys 525 520 Glu Glu Glu Lys Lys Asp Glu Glu Glu Lys Asp Glu Lys Glu Asp Ser 540 535 Lys Glu Asn Thr Lys Glu Lys Asp Lys Ile Asp Gly Thr Ala Glu Glu

555 545 550 Thr Glu Glu Arg Glu Gln Ala Thr Pro Arg Gly Arg Lys Thr Ala Asn 570 565 Ser Gln Gly Arg Arg Lys Gly Arg Ile Thr Arg Ser Met Thr Asn Glu 585 580 Ala Ala Ala Ala Ser Ala Ala Ala Ala Ala Ala Thr Glu Glu Pro Pro 600 Pro Pro Leu Pro Pro Pro Glu Pro Ile Ser Thr Glu Pro Val Glu 615 Thr Ser Arg Trp Thr Glu Glu Glu Met Glu Val Ala Lys Lys Gly Leu 635 630 Val Glu His Gly Arg Asn Trp Ala Ala Ile Ala Lys Met Val Gly Thr 650 645 Lys Ser Glu Ala Gln Cys Lys Asn Phe Tyr Phe Asn Tyr Lys Arg Arg 665 His Asn Leu Asp Asn Leu Leu Gln Gln His Lys Gln Lys Thr Ser Arg 680 Lys Pro Arg Glu Glu Arg Asp Val Ser Gln Cys Glu Ser Val Ala Ser 700 695 Thr Val Ser Ala Gln Glu Asp Glu Asp Ile Glu Ala Ser Asn Glu Glu 710 715 Glu Asn Pro Glu Asp Ser Glu Val Glu Ala Val Lys Pro Ser Glu Asp 730 725 Ser Pro Glu Asn Ala Thr Ser Arg Gly Asn Thr Glu Pro Ala Val Glu 740 745 Leu Glu Pro Thr Thr Glu Thr Ala Pro Ser Thr Ser Pro Ser Leu Ala 765 760 755 Val Pro Ser Thr Lys Pro Ala Glu Asp Glu Ser Val Glu Thr Gln Val 775 Asn Asp Ser Ile Ser Ala Glu Thr Ala Glu Gln Met Asp Val Asp Gln 795 790 Gln Glu His Ser Ala Glu Glu Gly Ser Val Cys Asp Pro Pro Pro Ala 810 805 Thr Lys Ala Asp Ser Val Asp Val Glu Val Arg Val Pro Glu Asn His 825 820 Ala Ser Lys Val Glu Gly Asp Asn Thr Lys Glu Arg Asp Leu Asp Arg 840 Ala Ser Glu Lys Val Glu Pro Arg Asp Glu Asp Leu Val Val Ala Gln 855 Gln Ile Asn Ala Gln Arg Pro Glu Pro Gln Ser Asp Asn Asp Ser Ser 875 870 Ala Thr Cys Ser Ala Asp Glu Asp Val Asp Gly Glu Pro Glu Arg Gln 890 885 Arg Met Phe Pro Met Asp Ser Lys Pro Ser Leu Leu Asn Pro Thr Gly 905 900 Ser Ile Leu Val Ser Ser Pro Leu Lys Pro Asn Pro Leu Asp Leu Pro 920 Gln Leu Gln His Arg Ala Ala Val Ile Pro Pro Met Val Ser Cys Thr 935 940 Pro Cys Asn Ile Pro Ile Gly Thr Pro Val Ser Gly Tyr Ala Leu Tyr 955 950 Gln Arg His Ile Lys Ala Met His Glu Ser Ala Leu Leu Glu Glu Gln 970 965 Arg Gln Arg Gln Glu Gln Ile Asp Leu Glu Cys Arg Ser Ser Thr Ser 985 980 Pro Cys Gly Thr Ser Lys Ser Pro Asn Arg Glu Trp Glu Val Leu Gln 1005 1000 Pro Ala Pro His Gln Leu Ile Thr Asn Leu Pro Glu Gly Val Arg Leu 1020 1015 Pro Thr Thr Arg Pro Thr Arg Pro Pro Pro Leu Ile Pro Ser Ser

1025	1030			1035		1040
Lys Thr Thr V	al Ala Ser ( 1045		1050			1055
Ile Ser Gln G	ly Thr Pro ( 060		1065		1070	)
Ser Tyr Thr G		1080			1085	
Gly Leu Pro A		1095		1100	)	
Ile Lys Gln G	1110			1115		1120
Gly Leu Leu V	1125		1130	)		1135
Gly Ala Ile G	140		1145		1150	)
Ile Ser Val G		1160			1165	
Thr Pro Ala I		1175		118	0	
Gly Ser Ile S	1190			1195		1200
Arg Glu Glu A	1205		1210	)		1215
Gly His Ile I	220		1225		123	U
Arg Ser Pro A		1240	)		1245	
Ser Val Glu (		1255		126	0	
Val Ser Ala H 1265	1270	)		1275		1280
Ser Pro His S	1285		129	0		1295
Met Gln Gly 5	1300		1305		131	U
Lys Tyr Pro 1		1320	)		1325	
Glu Gly Ala 1		1335		134	.0	
Lys Glu Met (	1350	)		1355		1360
Thr Gln Glu	1365		137	0		13/5
Ile Ile Glu	1380		1385		139	0
Asn Ser Gly 1395		1400	0		1405	
Gly Pro Ser 1410		1415		142	20	
Glu Asn Ile 1425	143	0		1435		1440
Gly Glu Thr	1445		145	0		1455
Ser Val Leu	1460		1465		147	70
Pro Gly Ile 1475		148	0		1485	
Gln Asn Thr 1490		1495		15	00	
Val Thr Ile	Pro Pro Asn	Lys Ser	Thr Asr	n His Gl	u Arg Lys	s ser Illi

1505	1510		1515		1520
1505 Leu Thr Pro Thr	Gln Ara Gl	n Ser Ile l		Ser Pro V	
	1525		1530	Ţ	.535
Gly Val Asp Pro	Val Val Se	er His Ser 1 1545	Pro Phe Asp	Pro His E 1550	lis Arg
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lie Pro Gly Inc	1685	m Pmc 110	1690		1695
Ser Met Ser Pro	Gly His P	ro Thr His	Leu Ala Ala	Ala Ala	Ser Ala
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Val Asp Ala Ala	1830 a Ala	ara Pro Gin	1835	1 331 -72	1840
Glu Ser Lys Hi	s Glu Ala A 1845	Ala Arg Leu	Glu Glu As: 1850	n Leu Arg	Ser Arg 1855
Ser Ala Ala Va	l Ser Glu G	Gln Gln Gln 186	Leu Glu Gl	n Lys Thr 1870	Leu Glu
18 Val Glu Lys Ar	g Ser Val G	ln Cys Leu	Tyr Thr Se		
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Ser Gly Lys Pr 1890	1	L895	19	00	
Lys Asp Lys Gl	1910		1915		1920
Thr Arg Gly Ly	s Thr Thr 1	Ile Thr Ala	Ala Asn Ph 1930	e Ile Asp	Val Ile 1935
Ile Thr Arg Gl	n Ile Ala S 40	Ser Asp Lys 194	Asp Ala Ar	g Glu Arg 1950	Gly Ser
Gln Ser Ser As	p Ser Ser S			s Arg Tyr	Glu Thr
1955		1960		1965	
Pro Ser Asp Al 1970	:	1975	19	80	
Pro Gln Glu Ly	s Leu Gln '	Thr Tyr Gln	Pro Glu Va	l Val Lys	Ala Asn

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1985				1990	)			`	1995	5				2000
Gln Ala	Glu	Asn	Asp 2009		Thr	Arg		Tyr 2010		Gly	Pro	Leu	His 2019	
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Ser Ser	Gln 2035	Ala		Gly		Gly 2040		Val	Pro	Arg	Thr 2045		Arg	Leu
Ile Thr		Ala	Asp	His		Cys		Ile	Ile	Thr 2060		Asp	Phe	Ala
Arg Asn 2065	Gln	Val	Ser	Ser 2070		Thr	Pro	Gln	Gln 2075		Pro	Thr	Ser	Thr 2080
Phe Gln	Asn	Ser	Pro 2085		Ala	Leu	Val	Ser 2090		Pro	Val	Arg	Thr 2099	_
Thr Ser	Asn	Arg 2100	_	Ser	Pro	Glu	Ser 2105		Ala	Gln	Ser	Val 2110		His
Gln Arg	2115	5		_		2120	)				2125	5		
Arg Gly 2130		Arg	Pro	Gly	Lys 2139		Pro	Glu	Arg	Ser 2140		Val	Ser	Ser
Glu Pro 2145	Tyr	Glu	Pro	Ile 2150		Pro	Pro	Gln	Val 2155		Val	Val	His	Glu 2160
Lys Gln			2165	5				2170	)				2179	5
Glu Gln	_	2180	)		_		2185	5				2190	)	
Ser Phe	2195	5	_			2200	)				2205	5		
Lys Gln 2210	)			_	2215	5				2220	)			
Asp Met 2225				2230	)	_			2235	5				2240
Val Thr			2245	5				2250	)				2255	5
Pro Ala		2260	)				2265	5				2270	)	
Gly Ser	2275	5				2280	)				2285	5		
Pro Met 2290	) _				2295	5				2300	)			
Gly Glu 2305				2310	)				2315	5				2320
Val Cys			2325	5				2330	)				2335	5
Ser Pro		2340	)				2345	5				2350	)	
Val Ser	2355	5				2360	)				2365	5		
Trp Ala	) "				2375	5				2380	)			
Tyr Asn 2385				2390	)				2395	5				2400
Ile Ala	-		2405	5				2410	)				2415	5
Asn Arg		2420	)				A1a 2425		ьeu	ьeu	ser	A1a 2430		Tyr
Glu Thr	ьец 2435		Авр	261	Asp	Asp 2440	)							